Volume 9, Issue 7 (2020) pp. 3916 - 3948



Anatomical study of Dendrobium (Orchidaceae) of Nepal

Baba Maiya Pradhan^{1*} and Devendra M. Bajracharya²

Department of Botany, Patan Multiple Campus, Tribhuvan University, Nepal.

Abstract: *Dendrobium* Swartz is one of the largest and most problematic genus of subtribe Dendrobiinae, tribe Malaxideae of family Orchidaceae. Twenty-six species of *Dendrobium* investigated are characterized by the variations in anatomical characters of leaves, stems, stomata and stegmata. These anatomical characters are taxonomically significant and used as distinguishing characters for delimiting taxa at infrageneric level. An artificial dichotomous key based on anatomical characters of leaves and stems for the identification of Dendrobium species of Nepal is provided. The cluster analysis based on anatomical with micromorphological characters is carried out with the aim of investigating interrelationships among the species of *Dendrobium* present in Nepal.

Keywords: Dendrobium; Orchidaceae; Anatomy; Stomata; Stegmata; Cluster analysis

Introduction

Dendrobium is a dominant genus of Orchidaceae and was established by Olof Swartz in 1799. Dendrobium moniliforme (L.) Sw. is the type for the genus Dendrobium (Holttum, Brieger and Cribb 1979). Species of Dendrobium are mostly epiphytes with some are lithophytes. Dendrobium Swartz comes under subtribe Dendrobiinae and it consists of about 1450 species worldwide (Schuiteman, 2014). Rajbhandari et al., (1999) reported that largest number of epiphytic orchids are found in the lower temperate regions of Central and Eastern Nepal, The Subtribe Dendrobiinae is represented by genera Dendrobium, Epigeneium and Flickingeria (Dressler, 1993). But according to update classification of Orchidaceae (Chase et al., 2015) two genera Epigeneium and Flickingeria were merged into single genus Dendrobium. The distinguishing characters of the genus Dendrobium are presence of cane-like stem with pseudo-bulb covered with or without sheathing leaf bases, distichous or duplicate leaves, spiral flowers with basally jointed lips with prominent spurs formed by the column foot and naked pollinia.

Although morphological and floristic studies of Orchidaceae have been carried out extensively, anatomical investigations are yet inadequate on orchids from Nepal. Only a few anatomical characters have been explored by some of the scientists as a supplementary portion of the revision of respective genera (Coelogyne, Dendrobium, Eria, Gastrochilus, Oberonia and Pholidota) in Nepal. In subtribe Dendrobiinae, only certain species have been studied anatomically as a part of a broad investigation (Morris et al., 1996). Kaushik (1983) explored the ecological and anatomical marvels of the Himalayan Orchids and discussed the impact of anatomy on the classification of the family and formulated the phylogenetic classification based on the morpho-anatomical characters. Morris et al., (1996) studied the vegetative anatomy and systematic of subtribe Dendrobiinae. They found that the anatomy of plants reflects a high degree of morphological diversity and many of the anatomical characters appear to be homoplasous. They also recommended that a more detailed understanding of the phylogeny of subtribe Dendrobiinae will require the characters from morphology, micromorphology, anatomy and DNA studies. Carlsward et al., (1997) studied the comparative leaf anatomy and systematic in Dendrobium, Sections Aporum and Rhizobium and concluded that two sections of Dendrobium are hypothesized to be sister taxa because of synapomorphies in their foliar anatomy. The cladistic analysis performed with various anatomical characters of the leaf demonstrates that both groups are monophyletic. Anatomical characterization helps in the identification of plant taxa when morphological characters are similar. It also helps in understanding the physiological processes and phylogenetic relationships among plant species (Liu and Zhu, 2011).

Micromorphological study on leaf epidermal surface had been widely used in taxonomic and phylogenetic studies. Singh and Singh (1974) studied the organization of stomatal complex in some Orchidaceae and concluded that the presence of tetracytic stomata was considered significant in evolutionary relationships of the family. Kaushik (1983) and Rasmussen (1987) studied the ecological significance of dermal characters. Khasim and Mohana Rao (1990) studied anatomy in relation to taxonomy in some members of Epidendrodoideae. Vij et al., (1991) studied the epidermal features of Indian orchids for taxonomic and ecological implications. Yukawa et al., (1992) reported the existence of two types of stomatal shape in the genus Dendrobium and its systematic significance. Arditti (1992) suggested that the shape of subsidiary cells had been used to distinguish the stomatal complex of Orchidaceae into several types. In concerning about Himalayan species, Shakya (1999) classified eight types of stomatal complex according to the arrangement of subsidiary cells among the twelve species of the genus Oberonia. Bajracharya (2003) observed seven types of stomata within the Himalayan genus Eria. Dangol (2006) studied the anatomical structures of subtribe Coelogyninae.

Stegmata or silica cell in orchids was reported and illustrated by Link (1849) for the first time but described them as projections from the sclerenchyma. The stegmata are restricted to the monocotyledon families Orchidaceae, Palmae, Maranthaceae, Musaceae, and with some doubt Juncaceae and not found in dicotyledons (Kohl, 1889). He distinguished two main kinds of stegmata in orchids one with conical bodies and other with spherical bodies. Stegmata with conical bodies are widespread within the family and those with spherical bodies are found in vandoid orchids and *Dendrobium*. Although the distribution of stegmata in orchids seems to be of systematic value and phylogenetic interest, little attention has been paid to them since then. Moller and Rasmussen (1984) studied the distribution of stegmata in 130 species representing 105 genera in Orchidales. They reported that the stegmata bearing silica bodies occur in longitudinal files lining fibre bundles and vascular bundle sheaths of rhizomes, stems and especially in the leaves and rarely in roots. Dressler (1993) reported that in most orchids the silica cells are roughly conical in shape but they are spherical and rather lumpy in the Vandae, Eriinae, Podochilinae and Dendrobiinae. Carlsward et al., (1997) reported that stegmata containing rough-surfaced spherical bodies are found associated with phloic sclerenchyma of the larger bundles of Dendrobiun leonis, D. anceps, D. brevimentum, D. aloifolium, D. distichum, D. indivisum, D. mannii, D. rosellum and D. nathaniele and they are absent along vascular bundle sclerenchyma in D. acinaciforme. Bajracharya (2003) studied the stegmata on leaf surface of 28 species of Eria. He reported that the study of stegmata reveal valuable characters that had been used in delimiting the taxa Eria at species level.

Presently 29 species of Dendrobium are reported from Nepal Himalaya. Out of 29 species, 26 species of Dendrobium were studied for anatomical structures. Remaining 3 species (D. darjeelingense Pradhan, D. chryseum Rolfe and D. plicatile Lindley) were excluded from anatomical study because these species could not be collected from field visits. This may be due to extinction of orchids from their natural habitats because of their medicinal values, deforestation or landslides. In concerning about D. darjeelingense it was known that this species was destroyed due to forest firing. This study will provide the taxonomic significance, relationships and variations among different species of Dendrobium of Nepal.

Materials and Methods

Study sites and sample collection

In this study, the plant materials were collected from West, Central and East Nepal on the basis of those regions mentioned in the herbaria and also from the available literature. The collected specimens were preserved in FAA (Formalin 0.5 parts, glacial acetic acid 0.5 parts, 70% ethanol 9 parts) and stored in 70% ethanol for anatomical studies. These techniques involve killing and fixing the tissues in FAA and then storing in 70 % ethanol before sectioning.

Anatomical study

Anatomical studies of the different parts of the plant body such as leaves and stems or pseudobulbs were studied on the collected materials by two methods *viz.*, Free hand sections and Microtomy.

In Free hand sections method, the plant parts were hand sectioned into 25-30 µm with the help of sharp blades and dehydrated with different alcohol series. Double staining was done by Safraninfast green combination (Johansen, 1940) and the sections were mounted in DPX. The different anatomical characters were observed and measured using compound light microscopes (Olympus) fitted with ocular micrometer scale. Microscopic photography was carried out of the observed specimens. In Microtomy method, sections were prepared using standard micro-technical methods for anatomy by using a rotary microtome (Yorco Spencer type) as described by Cutler (1978). For microtome sections, leaf and stem parts of specimens were dehydrated in alcohol and xylene series, infiltrated and embedded in paraffin wax (melting point 60-62º.c). The embedded materials in the block of wax were sectioned with a rotary microtome at a thickness of 15-30 µm. Staining, measurement and photography were same as in free hand sections method.

Micromorphological study Stomata

The leaf material for epidermal peeling was prepared by immersing in Potassium hydroxide solution (5%) overnight, rinsed with deionized water and again immersed in fresh solution of potassium hydroxide solution for few hours, then washed with water and treated with glacial acetic acid for 2-5 minutes (Carpenter 2005; with some modification). The peeled epidermal layer was then treated with Sodium hypochlorite (4%) until the tissue got discolored. It was then washed with water for several times and stained in Safranin (1%) and mounted in glycerin. The leaf epidermis was observed under light microscope. The size of stomata and stomatal pore of each specimen were measured by using the standardized ocular micrometer scale. The microscopic photography was carried out. The stomatal distribution on the both surfaces were observed and noted. The terminologies used for stomatal complex types were followed those of Patel (1979). The data were stored in the tabulated form. Stomatal index and stomatal frequencies were calculated using the formula given by Salisbury (1928).

$SI = S / E + S \times 100,$

Where, SI = Stomata Index, E = Average number of epidermal cells per unit area or microscopic field, S = Average number of stomata in microscopic field.

SF=S/A per mm²

Where, SF= Stomata frequency, A= Area of microscopic field.

$A = \pi r^2$

Where, $\pi = 3.14$, r = radius of the microscopic field.

Stegmata

Stegmata were studied on the leaf surfaces of specimen by peeling the epidermal layer, stained in Safranin (1%) and mounted on glycerin. The leaf epidermis was observed under light microscope and microscopic photography was carried out.

Cluster analysis

In the present study, the character and character states based on collected live specimens and herbarium specimens were described to carry out the cluster analysis. Altogether 60 characters (anatomical with micromorphological) were considered to perform the cluster analysis.

Data matrix

Altogether 60 variable anatomical characters were scored as multistate characters. The multistate characters were coded as 0-4. The complete data matrix of characters and character states are presented in Table 1. The data matrix of different species is presented in Table 2.

Data analysis

The cluster analysis was performed on the anatomical with micromorphological data with pvclust (Suzuki and Shimodaira, 2011), an R

package (R version 3.1.0, 2014). The pvclust is an implementation of bootstrap analysis on a statistical software R for hierarchical clustering with *p*-values. The pvclust calculates probability values (p-values) for each cluster during bootstrap resampling techniques. In pvclust, two types of *p*-values are available: Approximately Unbiased (AU) p-value and Boots-trap Probability (BP) *p*-value. Multiple bootstrap resampling is used for the calculation of AU *p*value which has superiority in bias over BP value calculated by normal bootstrap resampling. R-pvclust analysis was performed on anatomical data with 10000 bootstrapping to get the stronger cluster supported by data. The average cluster analysis with correlation distance was used to analyze the data set.

Results

Anatomy with micromorphology Leaf Surface

Epidermis: cells vary from square to rectangular to polygonal in shape. Rectangular to polygonal epidermal cells are present in species of all the species of Dendrobium except in D. bicameratum, D. chrysanthum, D. crepidatum, D. denneanum, D. densiflorum, D. denudans, D. fuscescens, D. fugax and D. porphyrochilum are foundsquare to polygonal epidermal cells are present.

Stomata: present on abaxial surface only in all species of *Dendrobium*. a, b and c tetra-monocyclic stomata are present in all species of *Dendrobium* except in *D. anceps* (eupara twi-monocyclic with hexa-monocyclic stomata) and *D. fugax* (eupara twi-monocyclic stomata).

Three types of stomata i.e. elliptical, circular and sub-orbicular are present in all species of *Dendrobium*. Largest stoma was found in *D*. *formosum* and smallest in *D*. *fuscescens*. Largest stomatal pore was found in *D*. *nobile* and smallest in *D*. *porphyrochilum*.

Stegmata: with roughsurfaced spherical silica bodies are present all the species of Dendrobiumand not present in *D. densiflorum*, *D. formosum*, *D. longicornu* and *D. polyanthum* (Table 3, Plates 1, 2, 3, 4, 5, 6, 7, 8).

Leaf (T.S.)

Cuticle: Thin cuticles are present in *D. aphyllum, D. crepidatum, D. denudans, D. eriiflorum, D. fimbriatum,D. formosum, D. gibsonii, D. heterocarpum, D. longicornu, D. monticola, D. moschatum, D. nobile, D. polyanthum, D. porphyrochilum* and *D. transparens* whereas thick cuticles are present in *D. amoenum, D. amplum, D. anceps, D. bicameratum, D. chrysanthum, D. denneanum, D. densiflorum, D. fuscescens, D. fugax, D. moniliforme* and *D. rotundatum.* Cuticle is intermittently domed in *D. anceps.*

Epidermis: cells vary from square to rectangular to hexagonal to polygonal. The adaxial cells are larger than abaxial cells.

Hypodermis: single layer of rounded small cells is present abaxially in *D. anceps* and adaxially in *D. densiflorum* and absent in other species.

Fibre bundles: present in one series both on adaxial and abaxial sides subtending the hypodermis in *D. anceps* and absent in other species.

Mesophyll: homogenous and heterogenous. Heterogenous mesophylls are present in *D*. *amplum* and *D*. *rotundatum* whereas homogenous mesophylls are present in remaining species. Cells are thin walled, columnar to oval to polygonal. Cell layers range from 6 in *D*. *monticola* to 18 in *D*. *anceps* in homogenous mesophylls. In heterogenous mesophylls, palisade layers 2 in *D*. *amplum* and *D*. *rotundatum* whereas spongy layers 7 in *D*. *amplum* and 8 in *D*. *rotundatum*.

Vascular bundle: conjoint, collateral and closed bundles are arranged in one row with a large bundle at midrib. Midrib vascular bundle is oval to oblong to flask shaped to circular to conical. U-shaped fibre caps are present at xylem poles and C-shaped fibre caps at phloem poles of midrib vascular bundles. In case of *D. moschatum*, and *D. densiflorum* dome-shaped fibre caps are present at xylem poles.

S.No.	Character	Character states
0	Stomatal shape	(0) elliptic; (1) circular; (2) sub-orbicular
1	Stomatal type	(0) a-tetra-monocyclic; (1) b-tetra-monocyclic; (2) c-tetra-monocyclic;(3)eupara-twi-monocyclic;(4) eupara-twi-monocyclic+hexa-monocyclic
2	Stomata length, μm	(0) < 33; (1) 33 - 37; (2) > 37
3	Stomata width, µm	(0) < 26; (1) 26 - 28; (2) > 28
4	Stomatal pore length, μm	(0) < 16; (1) 16 - 18; (2) > 18
5	Stomatal pore width, µm	(0) < 7; (1) 7 - 9.99; (2) > 9.99
6	Stomatal frequency, mm ²	(0) < 80; (1) 80-120; (2) > 120
7	Stomatal index, %	(0) < 7; (1) 7 - 10; (2) > 10 %
8	Leaf epidermal cell type	(0) parenchyma; (1) sclerenchyma
9	Stegmata	(0) absent; (1) present; (2) indistinct
10	Cuticle layer of leaf	(0) thick; (1) thin
11	Adaxial leaf epidermal cell width (largest), μm	(0) < 46; (1) 46 - 67; (2) > 67
12	Adaxial leaf epidermal cell length (largest), μm	(0) < 28; (1) 28 - 40; (2) > 40
13	Abaxial leaf epidermal cell width (largest), μm	(0) < 28; (1) 28 - 42; (2) > 42
14	Abaxial leaf epidermal cell length (largest), μm	(0) < 18; (1) 18 - 24; (2) > 24
15	Leaf hypodermis position	(0) absent; (1) adaxial; (2) abaxial
16	Leaf hypodermal layer	(0) absent; (1) 1
17	Mesophyll layer type	(0) homogenous; (1) heterogenous
18	Homogenous mesophyll layers	(0) absent; (1) \leq 7; (3) 8 - 10
19	Heterogenous mesophyll spongy layers	(0) absent; (1) 7; (2) 8
20	Water storage cells in mesophyll	(0) absent; (1) present
21	Calcium oxalate crystals in mesophyll	(0) absent; (1) present
22	Midrib vascular bundle shape	(0) oval; (1) flask-shaped; (2) conical; (3) oblong; (4) circular
23	Midrib vascular bundle length, μm	(0) < 140; (1) 140 - 200; (2) > 200
24	Midrib vascular bundle width, µm	(0) < 120; (1) 120 - 170; (2) > 170
25	Fibre bundle	(0) absent; (1) present
26	Major vascular bundle at leaf pole	(0) absent; (1) present
27	Fibre cap on midrib bundle	(0) absent; (1) present
28	Xylem fibre cap layers in midrib bundle	(0) absent; (1) < 4; (2) 4 - 6; (3) > 6
29	Phloem fibre cap layers in midrib bundle	(0) absent; (1) < 4; (2) 4 - 5; (3) > 5

Table 1. Characters and character states used in Cluster analysis of *Dendrobium*

30	Xylem fibre cap length, μm	(0) absent; (1) < 30; (2) 30 - 50; (3) > 50
31	Xylem fibre cap width, μm	(0) absent; (1) < 120; (2) 120 - 150; (3) > 150
32	Phloem fibre cap length, μm	(0) absent; (1) < 27; (2) $27 - 37$; (3) > 37
33	Phloem fibre cap width, μm	(0) absent; (1) < 130 (2) 130 - 175; (3) > 175
34	Xylem length (largest), μm	(0 < 19; (1) 19 - 25; (2) > 25
35	Xylem width (largest), μm	(0) < 10; (1) 10 - 17; (2) > 17
36	Phloem length (largest), μm	(0) < 6; (1) 6 - 9; (2) > 9
37	Phloem width (largest), μm	(0) < 4.0; (1) 4.0 - 6.5; (2) > 6.5
20	Stom anidarmal call change	(0) rectangular to polygonal; (1) rectangular to square;
30	Stem epidermai cen snape	(2) rectangular to angular; (3) rectangular; (4) barrel-shaped
39	Stem epidermal cell width (largest), μm	(0) < 18; (1) 18 - 25; (2) > 25
40	Stem epidermal cell length (largest), μm	(0) < 12; (1) 12 - 16; (2) > 16
41	Stem hypodermal layer	(0) absent; (1) 1; (2) 2 - 3; (3) 4 - 5
40	Stem hune dormal call shane	(0) absent; (1) oblong-Polygonal; (2) rectangular-polygonal;
42	Stem nypodermar cen snape	(3) polygonal; (4) oval-polygonal
43	Stem vascular bundle shape	(0) oval; (1) elongate
44	Stem vascular bundle no.	(0) $60; (1) 60 - 70; (2) > 70$
45	Stem vascular bundle length (smallest), µm	(0) < 100; (1) 100 - 130; (2) > 130
46	Stem vascular bundle width (smallest), µm	(0) < 80; (1) 80 - 90; (2) > 90
47	Stem vascular bundle length (largest), μm	(0) < 190; (1) 190 - 230; (2) > 230
48	Stem vascular bundle width (largest), μm	(0) < 100; (1) 100 - 130; (2) > 130
49	Fibre cap in vascular bundle of stem	(0) absent; (1) present
50	Xylem fibre cap length (largest), μm	(0) absent; (1) \leq 45; (2) > 45
51	Xylem fibre cap width (largest) μm,	(0) absent; (1) ≤ 100 ; (2) > 100
52	Phloem fibre cap length (largest), µm	(0) absent; (1) < 30 ; (2) 30 – 37; (3) > 37
53	Phloem fibre cap width (largest), µm	(0) absent; (1) < 80; (2) 80 - 100; (3) > 100
54	Xylem length (largest), μm	(0) < 18; (1) 18 - 27; (2) > 27
55	Xylem width (largest), μm	(0) < 15; (1) 15 - 20; (2) > 20
56	Phloem length (largest), μm	(0) < 7; (1) 7 - 10; (2) > 10
57	Phloem width (largest), μm	(0) < 5; (1) 5 - 8; (2) > 8
58	Stegmata in vascular bundle of stem	(0) absent; (1) present
59	Raphide bundles in stem	(0) absent; (1) present.

	Taxa									C]hai	act	ers	anc	l ch	ara	cte	r sta	ates	wi	th c	od	es								
S. N.	Dendrohium											1	1	1	1	1	1	1	1	1	1	1	2	2	2	2	2	2	2	2	2
	Denurootum	0	1	2	3	4	5	6	7	8	9	0	1	2	3	4	5	6	7	8	9	0	1	2	3	4	5	6	7	8	9
1	D. amoenum	0	1	0	0	0	0	0	0	0	1	1	2	1	2	1	0	0	0	1	0	0	1	0	1	1	0	0	1	2	1
2	D. amplum	2	1	1	2	1	0	1	0	1	1	0	1	1	2	1	0	0	1	0	1	0	0	1	1	1	0	0	1	2	3
3	D. anceps	0	4	0	0	0	0	0	0	0	2	0	0	0	0	0	2	1	0	3	0	1	1	4	0	1	1	1	1	1	1
4	D. aphyllum	0	0	1	0	1	1	0	1	0	1	1	2	2	2	2	0	0	0	2	0	1	0	0	0	0	0	0	1	1	1
;	D. bicameratum	0	0	1	2	2	1	0	1	0	2	0	1	1	1	0	0	0	0	2	0	0	0	0	0	0	0	0	1	1	1
5	D. chrysanthum	1	0	1	0	1	0	0	0	0	1	1	2	0	1	1	0	0	0	2	0	0	0	0	2	2	0	0	1	3	2
7	D. crepidatum	0	1	0	1	0	1	2	2	0	1	1	0	2	0	2	0	0	0	2	0	0	1	2	2	1	0	0	1	2	3
3	D. denneanum	1	0	0	1	1	1	2	1	1	1	0	0	0	0	0	0	0	0	2	0	0	1	2	2	2	0	0	1	2	2
)	D. densiflorum	0	0	1	1	2	2	0	0	1	0	0	0	0	0	1	1	1	0	3	0	0	1	2	2	2	0	0	1	3	3
.0	D. denudans	0	0	2	1	2	1	1	2	0	2	1	0	0	1	0	0	0	0	1	0	0	1	3	0	0	0	0	1	1	1
1	D. eriiflorum	0	2	2	0	2	2	2	2	0	2	1	0	2	0	2	0	0	0	1	0	0	1	0	0	0	0	0	1	2	2
12	D. fimbriatum	1	0	1	2	1	1	2	2	1	1	1	0	0	0	0	0	0	0	1	0	0	0	0	1	1	0	0	1	2	2
13	D. formosum	1	2	2	2	2	0	0	2	1	0	1	2	2	2	2	0	0	0	2	0	0	1	0	2	2	0	0	1	3	2
14	D. fugax	1	3	0	2	0	2	2	2	1	2	0	0	0	0	0	0	0	0	3	0	0	1	1	2	1	0	0	1	3	3
15	D. fuscescens	1	0	0	0	0	0	0	0	1	1	0	0	0	1	1	0	0	1	0	1	0	0	1	2	1	0	0	1	2	3
16	D. gibsonii	0	0	1	0	2	0	0	1	0	1	1	1	1	1	0	0	0	0	2	0	0	1	0	2	2	0	0	1	2	2
17	D. heterocarpum	1	0	2	2	1	2	1	2	0	1	1	2	2	1	2	0	0	0	1	0	0	1	0	1	0	0	0	1	2	1
18	D. longicornu	0	0	0	0	0	0	2	2	0	0	1	1	2	2	2	0	0	0	2	0	0	1	4	1	1	0	0	1	2	2
19	D. moniliforme	1	0	2	2	1	0	2	2	0	2	0	1	2	0	1	0	0	0	2	0	0	1	0	0	1	0	0	1	1	2
20	D. monticola	0	0	2	1	2	1	2	2	0	2	1	1	1	0	1	0	0	0	1	0	0	0	0	0	0	0	0	1	1	1
21	D. moschatum	1	0	0	1	1	1	0	0	0	1	1	1	0	0	0	0	0	0	3	0	1	1	2	2	2	0	0	1	3	3
22	D. nobile	0	2	2	2	2	2	1	2	0	1	1	2	1	2	2	0	0	0	2	0	1	1	0	1	1	0	0	1	1	1
23	D. polyanthum	0	1	0	1	1	0	0	1	0	0	1	0	0	1	0	0	0	0	2	0	1	0	4	2	2	0	0	1	3	2
24	D. porphyrochilum	0	0	1	0	0	0	0	1	0	1	1	2	1	1	1	0	0	0	1	0	1	1	3	0	0	0	0	0	0	0
25	D. rotundatum	2	2	0	0	0	0	1	1	1	2	0	1	1	1	2	0	0	1	0	2	1	1	1	1	1	0	0	1	2	3
26	D. transparens	0	1	1	1	1	0	2	2	0	1	1	1	2	2	1	0	0	0	2	0	0	1	4	1	1	0	0	1	2	2

Table 2: Data matrix for cluster analysis of Dendrobium

	Taxa									C	har	acte	ers	and	l ch	ara	cter	sta	tes	wit	h c	ode	es								
S. N.	Dendrohium	2	3	3	3	3	3	3	3	3	3	4	4	4	4	4	4	4	4	4	4	5	5	5	5	5	5	5	5	5	5
	Denarootam	0	1	2	3	4	5	6	7	8	9	0	1	2	3	4	5	6	7	8	9	0	1	2	3	4	5	6	7	8	9
1	D. amoenum	3	2	2	2	2	А	1	1	0	1	0	2	3	0	2	2	2	0	1	1	0	0	2	2	2	2	2	2	1	0
2	D. amplum	1	2	3	2	1	2	1	1	1	1	1	2	3	1	0	2	0	2	2	1	2	2	3	3	2	1	1	1	1	0
3	D. anceps	1	1	1	2	0	0	0	0	4	0	0	2	3	0	1	1	0	1	0	1	1	1	3	2	0	0	0	1	0	0
1	D. aphyllum	1	1	1	1	1	1	1	1	4	0	2	1	3	0	2	2	2	2	1	1	0	0	2	2	1	1	2	1	1	1
5	D. bicameratum	1	1	1	1	0	0	1	0	4	0	0	2	3	1	2	0	1	1	0	1	0	0	2	1	2	2	0	1	1	0
6	D. chrysanthum	2	3	2	3	1	2	2	2	4	1	1	2	4	1	1	1	1	2	2	1	0	0	2	2	2	2	1	1	0	1
7	D. crepidatum	2	2	3	2	1	1	1	1	0	2	0	1	1	0	0	1	1	1	0	1	0	0	1	1	1	1	1	2	0	0
3	D. denneanum	2	3	2	3	1	1	1	1	4	1	1	3	2	1	2	2	2	2	2	1	0	0	3	3	2	2	2	2	0	0
9	D. densiflorum	3	3	3	3	2	1	2	2	3	0	2	3	2	0	2	1	1	1	2	1	0	0	2	3	0	1	0	0	1	1
10	D. denudans	0	1	1	1	1	0	1	1	3	2	1	0	0	0	0	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0
11	D. eriiflorum	2	1	1	1	1	1	1	1	3	2	1	0	0	0	1	1	1	1	1	1	0	0	1	1	0	0	0	0	0	1
12	D. fimbriatum	2	1	2	1	1	1	1	1	4	0	0	3	2	0	1	2	2	1	2	1	0	0	3	3	2	0	2	2	0	0
13	D. formosum	3	3	2	3	2	2	2	2	0	2	2	2	3	0	1	2	2	2	2	1	0	0	3	3	2	0	2	2	1	0
14	D. fugax	3	3	2	3	2	1	2	2	4	1	2	2	3	1	2	1	1	1	2	1	0	0	2	2	1	1	0	1	1	0
15	D. fuscescens	3	1	3	3	1	1	2	2	1	2	2	2	3	0	2	0	1	2	1	1	2	2	1	1	1	1	1	1	0	0
16	D. gibsonii	3	3	3	3	2	2	2	2	2	0	1	2	3	0	0	1	2	2	2	1	0	0	3	2	2	1	2	2	0	1
17	D. heterocarpum	1	1	1	1	1	1	1	1	2	1	1	2	3	0	2	1	2	0	1	1	0	0	3	2	0	0	1	1	0	1
18	D. longicornu	2	3	2	1	1	1	1	2	0	1	1	2	3	1	1	0	1	0	1	1	0	0	3	2	1	1	2	2	1	1
19	D. moniliforme	1	1	1	2	0	0	0	0	0	1	1	2	3	1	0	2	2	2	2	1	0	0	1	1	1	1	2	2	0	1
20	D. monticola	1	1	1	1	0	0	1	1	3	2	0	0	0	1	0	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0
21	D. moschatum	3	3	3	3	2	2	2	2	4	0	2	2	3	0	2	2	2	0	2	1	1	1	2	3	0	0	0	0	0	1
22	D. nobile	2	3	1	3	1	2	2	2	0	2	1	2	3	0	2	1	1	1	2	1	0	0	2	2	1	1	0	1	0	0
23	D. polyanthum	3	3	3	2	1	1	1	1	0	2	2	1	3	1	0	0	0	2	2	1	0	0	2	3	2	2	2	2	1	1
24	D. porphyrochilum	0	0	0	0	0	0	0	0	0	2	2	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0
25	D. rotundatum	2	3	3	1	2	2	1	1	1	2	2	2	3	0	2	0	1	0	1	1	0	0	3	2	1	1	2	2	1	1
26	D. transparens	2	2	1	2	1	1	1	1	1	1	2	2	3	1	0	0	0	0	0	1	0	0	1	1	0	0	1	1	1	1

In D. fimbriatum xylem pole and phloem pole are completely surrounded by fibre cap. STEGMATA with rough surfaced spherical silica bodies are present along the outer margin of xylem and phloem caps in D. amoenum, D. chrysanthum, D. fugax and D. moschatum; along xylem cap in D. bicameratum; along phloem cap in D. aphyllum, D. crepidatum, D. denudans, D. fuscescens, D. gibsonii, D. nobile; outer margin of fibre bundle in D. anceps; indistinct in D. amplum, D. denneanum, D. densiflorum, D. fimbriatum, D. formosum, D. heterocarpum, D.longicornu, D. polyanthum, D. porphyrochilum, D. transparens; D. rotundatumand lacking in D. eriiflorum, D. moniliforme and D. monticola (Table 4, Plates 1,2, 3, 4, 5, 6, 7, 8).

Stem (T.S.)

Cuticle: thick and smooth in most species of *Dendrobium* but in *D. porphyrochilum* it is thin and smooth. Cuticle is intermittently in *D. anceps*.

Epidermis: cells vary from rectangular to square to polygonal to angular to barrel-shaped in shape. Cells are thin walled in *D. anceps, D. porphyrochilum* and thick walled in other species.

Hypodermis: one to two layers of polygonal parenchymatous cells are present in *D. anceps, D. aphyllum, D. crepidatum* and *D. polyanthum*. In remaining species, two to five layers of rectangular to oval to polygonal sclerenchymatous cells are present. Outer hypodermal cells are smaller than inner cells. Hypodermal cells are smaller than ground tissues.

Ground tissue: no differentiation of cortex, endodermis, pericycle and pith.A mass of thin walled parenchymatous cells with intercellular spaces are present in ground tissue. The cells are of two types: many, small, living, round to polygonal assimilatory parenchymatous cells and fewer, large, scattered, dead, water-storage cells with unpleated outlines in *D. anceps, D. crepidatum, D. densiflorum, D. fimbriatum,D. moschatum, D. nobile,D. polyanthum* and with pleated outlines in other species. Starch grains are present in parenchymatous cells in all species except *D. amoenum*, *D. anceps*, *D. denneanum*, *D. denudans*, *D. monticola* and raphide bundles are present in *D. aphyllum*, *D. chrysanthum*, *D. densiflorum*, *D. eriiflorum*, *D. gibsonii*, *D. heterocarpum*, *D. longicornu*, *D. moniliforme*, *D. moschatum*, *D. polyanthum*, *D. rotundatum* and *D. transparens*. Calcium oxalate crystals are present in *D. porphyrochilum*.

Vascular bundle: conjoint, collateral and closed bundles are scattered in the ground tissue. Number of vascular bundle ranges from 16 (D. polyanthum) to 118 (D. bicameratum). Vascular bundles are oval to elongate in shape. The inner vascular bundles are larger than outer bundles. Sclerenchymatous bundle sheaths are present surrounding vascular bundles in D. amoenum, D. anceps, D. chrysanthum, D. crepidatum, D. denudans, D. eriiflorum. D. fuscescens, D. heterocarpum, D. longicornu, D. porphyrochilum and only xylem in D. amplum, D. aphyllum, D. bicameratum, D. denneanum, D. densiflorum, D. fimbriatum, D. formosum, D. fugax, D. gibsonii, D. moniliforme, D. monticola, D. moschatumD. nobile, *D. polyanthum*, *D. rotundatum d D. transparens*. Fibre caps are present above phloem in D. amoenum, D. longicornu, D. monticola; surroundding xylem and phloem in D. anceps, D. fuscescens and surrounding phloem in remainning species.

Stegmata with rough surfaced spherical silica bodies are present along the outer margin of phloem cap in D. amoenum, D. amplum, D. aphyllum, D. bicameratum, D. densiflorum, D. formosum, D. fugax, D. longicornu, D. polyanthum, D. rotundatum, D. transparens; absent in D. eriiflorum, D. moniliforme, D. monticola and indistinct in remaining species (Table 5, Plates 1, 2, 3, 4, 5, 6, 7, 8). From the present study, the stomatal complex, stegmata and anatomical characters of leaves and stems were found to be taxonomically important for delimiting the taxa. Based on characters such as stomatal complex, stegmata and anatomical characters of leaves and stems, an anatomical key has been prepared to delimit the taxa within genus Dendrobium.

Table 3: Stomatal and Stegmatal characters in species of *Dendrobium*

S.No.	Taxa	Stomatal size with guard cells (l x w) µm	Stomatal pore size (l x w) μm	Stomatal Frequency mm²	Stomatal Index %	Shape of Stomata	Type of Stomata	Shape of Epidermal cells	Type of Epidermal cells	Stegmata
1	Dendrobium amoenum Wallich ex Lindley	29.97 x 23.31	13.32 x 4.99	76.92	5.81	Elliptic	b-tetra-monocyclic	Rect. to poly.	Paren.	Spherical
2	D. amplum Lindley	33.30 x 29.97	16.65 x 6.66	107.69	6.03	Sub- orbicular	b-tetra-monocyclic	Rect. to poly.	Scleren.	Spherical
3	D. anceps Swartz	29.97 x 25.30	13.32 x 6.66	46.15	3.75	Elliptic	eupara twi + hexa- monocyclic	Rect. to poly.	Paren.	Indistinct
4	D. aphyllum (Roxburgh) C.E.C. Fischer	33.30 x 23.31	16.65 x 9.99	46.15	7.31	Elliptic	a-tetra-monocyclic	Rect. to poly.	Paren.	Spherical
5	D. bicameratum Lindley	33.30 x 29.97	23.31 x 9.99	76.92	7.04	Elliptic	a-tetra-monocyclic	Squa. to poly.	Paren.	Indistinct
6	D. chrysanthum Wallich ex Lindley	33.30 x 23.31	16.65 x 6.66	76.92	5.68	Circular	a-tetra-monocyclic	Squa. to poly.	Paren.	Spherical
7	D. crepidatum Lindley & Paxton	32.63 x 26.64	15.32 x 7.32	130.77	13.04	Elliptic	b-tetra-monocyclic	Squa. to poly.	Paren.	Spherical
8	D. denneanum Kerr	26.64 x 26.64	16.65 x 9.99	123.07	7.34	Circular	a-tetra-monocyclic	Squa. to poly.	Scleren.	Spherical
9	D. densiflorum Wallich	33.30 x 26.64	26.64 x 13.32	76.92	6.76	Elliptic	a-tetra-monocyclic	Squa. to poly.	Scleren.	N.P
10	D. denudans D. Don	39.96 x 26.64	26.64 x 9.99	92.3	10.71	Elliptic	a-tetra-monocyclic	Squa. to poly.	Paren.	Indistinct
11	D. eriiflorum Griffith	38.62 x 25.97	20.64 x 10.65	123.07	10.66	Elliptic	c-tetra-monocyclic	Rect. to poly.	Paren.	Indistinct
12	D. fimbriatum Hooker	33.30 x 34.60	16.65 x 9.99	123.07	10.38	Circular	a-tetra-monocyclic	Rect. to poly.	Scleren.	Spherical
13	D. formosum Roxburgh ex Lindley	45.96 x 41.96	24.64 x 6.66	76.92	12.5	Circular	c-tetra-monocyclic	Rect. to poly.	Scleren.	N.P
14	D. fugax Reichenbach	29.97 x 29.97	13.32 x 13.32	184.61	10.17	Circular	eupara twi- monocyclic	Squa. to poly.	Scleren.	Indistinct
15	D. fuscescens Griffith	23.31 x 23.31	13.32 x 6.66	76.92	5.37	Circular	a-tetra-monocyclic	Squa. to poly.	Scleren.	Spherical
16	D. gibsonii Paxton	33.30 x 24.98	19.98 x 6.66	76.92	8.33	Elliptic	a-tetra-monocyclic	Rect. to poly.	Paren.	Spherical
17	D. heterocarpum Wallich ex Lindley	38.30 x 33.30	16.65 x 10.00	92.3	13.04	Circular	a-tetra-monocyclic	Rect. to poly.	Paren.	Spherical
18	D. longicornu Lindley	29.97 x 23.31	13.32 x 4.99	123.07	10.53	Elliptic	a-tetra-monocyclic	Rect. to poly.	Paren.	N.P
19	D. moniliforme (L.) Swartz	38.30 x 34.00	16.65 x 6.66	138.5	12.5	Circular	a-tetra-monocyclic	Rect. to poly.	Paren.	Indistinct
20	D. monticola Hunt & Summerhayes	39.96 x 26.64	23.31 x 9.99	200	10.83	Elliptic	a-tetra-monocyclic	Rect. to poly.	Paren.	Indistinct
21	D. moschatum (Buchanan-Hamilton) Swartz	27.97 x 26.64	16.65 x 9.99	76.92	5	Circular	a-tetra-monocyclic	Rect. to poly.	Paren.	Spherical
22	D. nobile Lindley	44.95 x 39.96	27.30 x 17.98	92.31	15.79	Elliptic	c-tetra-monocyclic	Rect. to poly.	Paren.	Spherical
23	D. polyanthum Wallich ex Lindley	32.63 x 26.64	16.65 x 4.99	76.92	7.94	Elliptic	b-tetra-monocyclic	Rect. to poly.	Paren.	N.P
24	D. porphyrochilum Lindley	36.63 x 19.98	13.32 x 3.33	76.92	7.14	Elliptic	a-tetra-monocyclic	Squa. to poly.	Paren.	Spherical
25	D. rotundatum (Lindley) Hooker	26.64 x 23.31	13.32 x 6.66	92.3	8.33	Sub- orbicular	c-tetra-monocyclic	Rect. to poly.	Scleren.	Indistinct
26	D. transparens Wallich ex Lindley	33.30 x 27.31	17.98 x 6.66	138.46	10.71	Elliptic	b-tetra-monocyclic	Rect. to poly.	Paren.	Spherical

Abbreviations used: N.P = Not present; Rect. = Rectangular; poly. = Polygonal; Squa. = Square; Paren = Parenchyma; Scleren. = Sclerenchyma.

]	Table 4: Anatomical	characte	ers of lea	af in speci	ies of <i>Der</i>	ıdrobium										
			I	Epidermal o Size (w	cell x l) μm	Hypode	ermis		Mesophy	711	Mid Va bun	scular dle	Fibre	cap size	Xylem	Phloem
S.No.	Таха	Cuticle layer	Shape	Adaxial	Abaxial	Position	Layer	Fibre bundle	Shape	Layer	Shape	Size (l x w) µm	Xylem cap (l x w) µm	Phloem cap (l x w) µm	size (l x w) µm	size (l x w) µm
1	<i>Dendrobium amoenum</i> Wallich ex Lindley	Thin	Rect. to Poly.	19.98- 69.93 x 19.98- 33.30	13.32- 43.29 x 9.99- 23.31	N.P	N.P	N.P	Colu. to Poly.	7	Oval	193.14 x 133.20	83.25 x 143.19	33.30 x 133.20	9.99 – 29.97 x 6.66 – 26.64	4.99-6.66 x 3.33- 4.99
2	D. amplum Lindley	Thick	Rect. to Poly.	19.98- 56.61 x 16.65- 29.97	16.65- 49.95 x 9.99- 19.98	N.P	N.P	N.P	Pali. Poly. to Colu. Spon.Oval to Poly.	Pali. 2 Spon. 7	Flask- shaped	199.80 x 143.19	16.65 x 139.86	46.62 x 166.50	6.66–23.31 x 4.99– 19.98	3.33-6.66 x 2.66- 4.99
3	D. anceps Swartz	Thick	Poly.	16.65– 36.63 x 6.66– 13.32	16.65- 23.31 x 6.66 - 13.32	Abaxial	1	Present	Oval to Poly.	18	Circular	139.06 x 136.53	23.31 x 119.88	26.64 x 139.86	6.66 - 9.99 x 4.99 - 9.99	3.33-4.99 x 2.66- 3.33
4	D. aphyllum (Roxburgh) C.E.C. Fischer	Thin	Rect. to Hexa.	46.62– 76.59 x 36.63– 49.95	23.31– 66.60 x 13.32– 29.97	N.P	N.P	N.P	Oval to Poly.	9	Oval	126.54 x 106.56	19.98 x 86.58	19.98 x 89.91	9.99–19.98 x 6.66 – 16.65	4.99–6.66 x 3.33– 4.99
5	D. bicameratum Lindley	Thick	Rect. to Poly.	13.32- 49.95 x 19.98- 29.97	13.32– 36.63 x 9.99– 13.32	N.P	N.P	N.P	Oval to Poly.	8	Oval	119.88 x 93.24	26.64 x 89.91	19.98 x 93.24	6.66 – 13.32 x 4.99 – 6.66	3.99–6.66 x 2.66– 3.33
6	D. chrysanthum Wallich ex Lindley	Thin	Rect. to Poly.	19.98- 73.26 x 16.65- 26.64	9.99– 36.63 x 9.99– 19.98	N.P	N.P	N.P	Rect.	8	Oval	219.79 x 209.78	49.95 x 186.48	36.63 x 183.15	9.9–19.98 x 8.32–19.98	6.66–9.99 x 3.33– 6.66
7	<i>D. crepidatum</i> Lindley & Paxton	Thin	Rect. to Poly.	19.98- 43.29 x 13.32- 43.29	8.32– 26.64 x 13.32– 26.64	N.P	N.P	N.P	Rect. to Poly.	10	Conical	233.10 x 166.50	43.29 x 126.54	53.28 x 173.16	13.32– 19.98 x 11.65– 13.32	4.99-8.32 x 3.33- 4.99

8	D. denneanum Kerr	Thick	Rect. to Poly.	19.98– 33.30 x 13.32– 16.65	16.65– 26.64 x 8.32– 9.99	N.P	N.P	N.P	Rect. to Poly.	8	Conical	216.45 x 189.81	49.95 x 166.50	33.30 x 176.49	6.66–19.98 x 4.99– 16.65	3.33–6.66 x 3.33– 4.99
9	D. densiflorum Wallich	Thick	Rect. to Poly.	23.31- 36.63 x 13.32- 23.31	16.65- 26.64 x 13.32- 19.98	Adaxial	1	N.P	Oval to Poly.	13	Conical	586.52 x 386.57	213.28 x 319.92	146.63 x 413.23	16.65– 29.97 x 6.66–16.65	4.99–9.99 x 3.33– 6.66
10	D. denudans D. Don	Thin	Rect. to Poly.	23.31- 43.29 x 9.99- 19.98	23.31– 29.97 x 9.99– 16.65	N.P	N.P	N.P	Rect. to Poly.	7	Oblong	126.54 x 53.28	23.31 x 53.28	23.31 x 56.61	13.32– 19.98 x 4.99–6.66	4.99-6.66 x 3.99- 4.99
11	D. eriiflorum Griffith	Thin	Rect. to Poly.	19.98– 43.29 x 26.64– 43.29	13.32– 23.31 x 19.98– 29.97	N.P	N.P	N.P	Rect. to Poly.	7	Oval	116.55 x 69.93	43.39 x 59.94	23.31 x 76.59	6.66–19.98 x 4.99– 13.32	3.33-6.66 x 2.66- 4.99
12	D. fimbriatum Hooker	Thin	Rect. to Poly.	16.65– 39.96 x 13.32– 26.64	9.99– 23.31 x 6.66– 16.65	N.P	N.P	N.P	Rect. to Poly.	7	Oval	169.83 x 123.21	43.29 x 116.55	33.30 x 129.80	9.99–19.98 x 9.99– 16.65	3.33-6.66 x 1.66- 4.99
13	D. formosum Roxburgh ex Lindley	Thin	Rect. to Poly.	36.63– 76.59 x 33.30– 46.62	19.98– 43.29 x 19.98– 33.30	N.P	N.P	N.P	Oval to Poly.	9	Oval	216.45 x 193.14	73.26 x 183.15	33.30 x 206.46	13.32– 29.97 x 6.66–19.98	3.33- 13.32 x 3.33-9.99
14	<i>D. fugax</i> Reichenbach	Thick	Rect. to Squa.	19.98– 33.30 x 9.99– 16.65	13.32– 23.31 x 9.99– 16.65	N.P	N.P	N.P	Oval to Poly.	15	Flask- shaped	223.11 x 166.50	56.61 x 166.50	36.63 x 199.80	9.99–26.64 x 9.99– 16.65	4.99-9.99 x 3.33- 6.66
15	D. fuscescens Griffith	Thick	Rect. to Poly.	23.31– 43.29 x 16.65– 26.64	16.65– 39.96 x 16.65– 23.31	N.P	N.P	N.P	Pali. Poly. to Oval Spon.Oval to Poly.	Pali. 2 Spon. 7	Flask- shaped	209.79 x 156.51	56.61 x 113.22	49.95 x 179.82	9.99–23.31 x 6.66– 16.65	4.99–9.99 x 4.99– 6.66

16	D. gibsonii Paxton	Thin	Rect. to Poly.	23.31– 66.60 x 13.32– 33.30	16.65– 33.30 x 13.32– 16.65	N.P	N.P	N.P	Rect. to Poly.	8	Oval	289.71 x 233.10	83.25 x 199.80	49.95 x 233.10	13.32– 29.97 x 6.66–23.31	4.99-9.99 x 3.33- 8.32
17	D. heterocarpum Wallich ex Lindley	Thin	Rect. to Poly.	26.64- 73.26 x 26.64- 53.28	9.99– 33.30 x 13.32– 26.64	N.P	N.P	N.P	Rect. to Poly.	7	Oval	149.85 x 116.55	19.98 x 99.90	19.98 x 86.58	6.66–19.98 x 4.99– 16.65	3.99-6.66 x 3.66- 4.99
18	D. longicornu Lindley	Thin	Rect. to Hexa.	23.31– 53.28 x 19.98– 46.62	19.98– 49.95 x 13.32– 33.30	N.P	N.P	N.P	Oval to Poly.	8	Circular	166.50 x 166.50	33.30 x 156.51	33.30 x 126.54	9.99–23.31 x 6.66– 16.65	4.99-6.66 x 3.33- 6.66
19	D. moniliforme (L.) Swartz	Thick	Rect. to Poly.	23.31- 56.61 x 36.63- 46.62	9.99 - 26.64 x 13.32- 19.98	N.P	N.P	N.P	Rect. to Poly.	8	Oval	119.88 x 133.20	26.64 x 106.56	23.31 x 133.20	9.99–10.65 x 4.99–6.66	3.99-5.99 x 2.66- 3.33
20	<i>D. monticola</i> Hunt & Summerhayes	Thin	Rect. to Poly.	13.32– 53.28 x 19.98– 33.30	13.32– 19.98 x 9.99– 19.98	N.P	N.P	N.P	Rect. to Poly.	6	Oval	99.90 x 66.60	23.31 x 53.28	13.32 x 66.60	6.66-9.99 x 3.33-8.32	4.99-6.66 x 3.33- 4.99
21	D. moschatum (Buchanan- Hamilton) Swartz	Thin	Rect. to Hexa.	26.64– 66.60 x 16.65– 26.64	16.65– 23.31 x 9.99– 16.65	N.P	N.P	N.P	Oval to Poly.	16	Conical	506.54 x 333.25	159.96 x 319.92	106.64 x 333.25	19.98- 39.96 x 6.66-26.64	4.99-9.99 x 3.33- 8.32
22	D. nobile Lindley	Thin	Rect. to Hexa.	36.63- 83.25 x 73.26- 99.90	16.65- 53.28 x 23.31- 46.62	N.P	N.P	N.P	Oval to Poly.	9	Oval	199.80 x 166.50	49.95 x 166.50	26.64 x 159.84	19.98– 23.31 x 9.99–19.98	6.66–9.99 x 4.99– 6.66
23	<i>D. polyanthum</i> Wallich ex Lindley	Thin	Rect. to Hexa.	16.65– 33.30 x 9.99– 26.64	13.32– 29.97 x 9.99– 13.32	N.P	N.P	N.P	Oval to Poly.	8	Circular	213.12 x 199.80	83.25 x 193.14	43.29 x 173.16	6.66–23.31 x 3.33–6.66	4.99-6.66 x 3.33- 4.99

24	D. porphyrochilum Lindley	Thin	Rect. to Hexa.	49.95- 73.26 x 19.98- 39.96	23.31- 39.96 x 16.65- 23.31	N.P	N.P	N.P	Oval to Poly.	6	Oblong	99.90 x 93.24	N.P	N.P	4.99–13.32 x 3.99–9.99	3.33-4.99 x 2.66- 3.33
25	D. rotundatum (Lindley) Hooker	Thick	Rect. to Poly.	23.31- 49.95 x 16.65- 33.30	9.99– 39.96 x 16.65– 26.64	N.P	N.P	N.P	Pali. Poly. to Colu. Spon.Oval to Poly.	Pali. 2 Spon. 8	Flask- shaped	183.15 x 133.20	39.96 x 193.14	53.28 x 126.54	13.32– 26.64 x 6.66–19.98	4.99-6.66 x 3.33- 4.99
26	D. <i>transparens</i> Wallich ex Lindley	Thin	Rect. to Hexa.	23.31– 66.60 x 36.63– 59.94	13.32- 43.29 x 13.32- 23.31	N.P	N.P	N.P	Oval to Poly.	8	Circular	149.85 x 146.52	33.30 x 149.85	26.64 x 146.52	13.32– 19.98 x 9.99–16.65	3.33-6.66 x 3.33- 4.99

Abbreviations used: N.P=Not Present; Rect.=Rectangular; Poly.=Polygonal; Squa.= Square; Hexa.=Hexagonal; Colu.=Columnar; Pali.=Palisade layer; Spon.=Spongy layer.

Table 5: Anatomical characters of stem in species of Dendrobium

			Epider	mal cell	Hypod	ermis	V	ascula	r bundle	Fibr	e cap size		Phloom
S.N.	Taxa	Cuticle	Shape	Size (w x l) μm	Shape	Layer	Shape	No.	Size (l x w) µm	Xylem cap (l x w) µm	Phloem cap (l x w) µm	Xylem size (l x w) μm	size (l x w) µm
1	<i>Dendrobium amoenum</i> Wallich ex Lindley	Thick	Rect. to Poly.	9.99–19.98 x 4.99-9.99	Poly.	2	Oval	76	143.19-183.15 x 93.24-113.22	N.P	26.64–33.30 x 83.25–96.57	9.99–29.97 x 6.66–23.31	6.66–16.65 x 4.99–9.99
2	D. amplum Lindley	Thick	Rect. to Squa.	9.99–19.98 x 9.99–13.32	Poly.	2	Elongate	50	133.20–333.00 x 66.60–149.85	43.29– 56.61 x 106.56– 133.20	33.30–99.90 x 83.25–116.55	11.65–26.64 x 9.99–16.65	3.33–9.99 x 3.33–6.66
3	D. anceps Swartz	Thick	Barr.	9.99–16.65 x 6.66-9.99	Poly.	2	Oval	64	116.55-199.80 x 73.26–99.90	16.65- 26.64 x 79.92- 86.58	19.98–49.95 x 73.26–99.90	13.32-16.65 x 4.99-13.32	4.99-6.66 x 3.33-6.66
4	<i>D. aphyllum</i> (Roxburgh) C.E.C. Fischer	Thick	Barr.	9.99-13.32 x 16.65-26.64	Poly.	1	Oval	82	133.20-236.43 x 103.23- 106.56	N.P	29.97–33.30 x 99.90–93.24	13.32–26.64 x 9.99–19.98	6.66–13.32 x 4.99–6.66
5	D. bicameratum Lindley	Thick	Barr.	9.99–13.32 x 6.66-9.99	Poly.	2 to 3	Elongate	118	93.24–199.80 x 83.25–93.24	N.P	16.65–33.30 x 49.95–66.60	13.32–33.30 x 6.66–26.64	4.99-6.66 x 3.33-6.66

6	D. chrysanthum Wallich ex Lindley	Thick	Barr.	13.32-19.98 x 9.99-13.32	Oval poly.	to	2	Elongate	70	103.23–266.40 x 83.25–133.20	N.P	16.65–33.30 x 83.25–96.57	9.99–33.30 x 9.99–29.97	4.99–9.99 x 3.33–6.66
7	D. crepidatum Lindley & Paxton	Thick	Rect. to Poly.	13.32–36.63 x 6.66-6.65	Oblo. Poly.	to	1	Oval	60	123.21–209.79 x 83.25–99.90	N.P	19.98–26.64 x 49.95–66.60	13.32–26.64 x 6.66–19.98	4.99–9.99 x 3.33–9.32
8	D. denneanum Kerr	Thick	Barr.	9.99–19.98 x 6.66-13.32	Rect. Poly.	to	4	Elongate	97	163.17–299.70 x 143.19– 149.85	N.P	39.96–49.95 x 116.55–119.88	43.29–49.95 x 23.31–23.31	9.99- 13.32 x 6.66-9.99
9	D. densiflorum Wallich	Thick	Rect.	6.66–9.99 x 16.65–16.65	Rect. Poly.	to	4 to 5	Oval	78	106.56–206.46 x 89.91–153.18	N.P	26.64-36.63 x 83.25-119.88	9.99-13.32 x 6.66-9.99	3.33–4.99 x 2.66–3.33
10	D. denudans D. Don	Thick	Rect.	13.32–26.64 x 6.66–13.32	N.P		N.P	Oval	50	66.60–119.88 x 56.61–76.59	N.P	16.65–16.65 x 49.95–56.6	9.99–13.32 x 6.66- 9.99	3.33–4.99 x 2.66–3.33
11	D. eriiflorum Griffith	Thick	Rect.	13.32–26.64 x 6.66–13.32	N.P		N.P	Oval	70	113.22–199.80 x 86.58- 126.54	N.P	19.98–29.97 x 49.95–66.60	13.32–16.65 x 9.99–13.32	3.33-6.66 x 3.33-4.99
12	D. fimbriatum Hooker	Thick	Barr.	13.32–16.65 x 9.99–11.65	Rect. Poly.	to	4 to 5	Oval	62	133.20-199.80 x 109.89- 133.20	N.P	39.96-43.29 x 106.56-123.21	26.64–49.95 x 9.99–13.32	6.66–16.65 x 4.99–9.99
13	<i>D. formosum</i> Roxburgh ex Lindley	Thick	Rect. to Poly.	16.65–39.96 x 9.99–16.65	Poly.		2	Oval	64	166.50-233.10 x 119.88- 153.18	N.P	33.30–53.28 x 89.91–109.89	26.64-49.95 x 9.99-13.32	6.66–16.65 x 4.99–9.99
14	D. fugax Reichenbach	Thick	Barr.	13.32–23.31 x 9.99–16.65	Poly.		2	Elongate	71	116.55-216.45 x 86.58 - 166.50	N.P	26.64–33.30 x 79.92 – 99.90	13.32-23.31 x 9.99-16.65	4.99-6.66 x 3.33-6.66
15	D. fuscescens Griffith	Thick	Rect. to Squa.	9.99–26.64 x 6.66–19.98	Poly.		2	Oval	83	99.90-243.09 x 89.91-116.55	16.65– 49.95 x 59.94– 116.55	16.65–26.64 x 59.94–66.60	6.66–23.31 x 6.66–16.65	3.33–9.99 x 3.33–6.66
16	D. gibsonii Paxton	Thick	Rect. to Angu.	13.32–16.65 x 6.66–13.32	Poly.		2	Oval	30	123.31–316.35 x 93.24-183.15	N.P	16.65–69.93 x 76.59–93.24	16.65–29.97 x 9.99–19.98	4.99–13.32 x 3.33–9.99
17	<i>D. heterocarpum</i> Wallich ex Lindley	Thick	Rect. to Angu.	13.32–19.98 x 6.66–13.32	Poly.		2	Oval	80	116.55–179.82 x 99.90–109.89	N.P	16.65–43.29 x 73.26–83.25	9.99–16.65 x 4.99–13.32	6.66–9.99 x 4.99–6.66
18	D. longicornu Lindley	Thick	Rect. to Poly.	13.32–19.98 x 6.66–13.32	Poly.		2	Elongate	68	76.59–183.15 x 89.91–119.88	N.P	16.65–46.62 x 79.92–83.25	9.99–26.64 x 4.99–19.98	4.99–11.65 x 3.33–9.99
19	D. moniliforme (L.) Swartz	Thick	Rect. to Poly.	13.32-23.31 x 6.66-13.32	Poly.		2	Elongate	55	199.80-233.10 x 113.22- 166.50	N.P	23.31–29.97 x 49.95–66.60	19.80–23.31 x 13.32–19.98	9.99-13.32 x 4.99-9.99
20	<i>D. monticola</i> Hunt & Summerhayes	Thick	Rect.	16.65–36.63 x 6.66–9.99	N.P		N.P	Elongate	20	66.60–89.91 x 49.95–53.28	N.P	13.32–16.65 x 43.29 – 49.95	6.66–13.32 x 4.99–9.99	4.99–6.66 x 3.33–4.99

21	D. moschatum (Buchanan-Hamilton) Swartz	Thick	Barr.	9.99–13.32 x 13.32–16.65	Poly.	3	Oval	78	133.20-166.50 x 113.22- 133.20	26.64- 43.29 x 66.60- 99.90	33.30-36.63 x 113.22-133.20	13.32-16.65 x 9.99-13.32	3.33-6.66 x 3.33-4.99
22	D. nobile Lindley	Thick	Rect. to Poly.	13.32–26.64 x 6.66–13.32	Poly.	2	Oval	82	116.55–216.45 x 86.58–166.50	N.P	26.64–33.30 x 79.92 – 99.90	13.32–19.98 x 9.99–16.65	4.99–6.66 x 3.33–6.66
23	<i>D. polyanthum</i> Wallich ex Lindley	Thick	Rect. to Poly.	19.98–39.96 x 13.32–16.65	Poly.	1	Elongate	16	99.90-289.71 x 49.95-133.20	N.P	33.30–36.30 x 63.27–119.88	9.99–39.96 x 6.66–23.31	4.99–16.65 x 3.33–13.32
24	D. porphyrochilum Lindley	Thin	Rect. to Poly.	29.97-46.62 x 19.98-23.31	N.P	N.P	Elongate	40	89.91–166.50 x 66.60– 83.25	N.P	N.P	9.99–16.65 x 6.66–9.99	6.66–8.32 x 3.33–6.66
25	<i>D. rotundatum</i> (Lindley) Hooker	Thick	Rect. to Squa.	16.65–29.97 x 9.99–16.65	Poly.	2	Oval	82	99.90–166.50 x 83.25–116.55	N.P	13.32–39.96 x 33.30–99.90	16.65–23.31 x 13.32–19.98	3.33–13.32 x 3.33–9.99
26	<i>D. transparens</i> Wallich ex Lindley	Thick	Rect. to Squa.	13.32–19.98 x 9.99–19.98	Poly.	2	Elongate	50	96.57–133.32 x 63.27–79.92	N.P	23.31–26.64 x 59.94–76.59	9.99–16.65 x 6.66–9.99	6.66–8.32 x 3.33–6.66

Abbreviations used: N.P = Not present; Rect. = Rectangular; Poly. = Polygonal; Angu. = Angular; Squa. = Square; Barr. = Barrel-shaped.





PLATE 1: D. longicornu A. Stomata, B. T.S. Leaf, C. T.S. Stem; D. aphyllum D., Stomata, E. T.S. Leaf, F. T.S. Stem; D. moschatum G. Stomata, H. T.S. Leaf, I. T.S. Stem; D. formosum J. Stomata, K. T.S. Leaf, L. T.S. Stem; D. rotundatum M. Stomata, N. T.S. Leaf, O. T.S. Stem. Stomata showing stoma (s), subsidiary cell (sc), epidermal cell (ec); T.S. Leaf showing cuticle (c), epidermis (ep), mesophyll cell (mc), xylem (x), phloem (p), fibre cap (fc), palisade layer (pl), water-storage cell (wc); T.S. Stem showing cuticle (c), epidermis (ep), leaf trace bundle (ltb), ground tissue (gt), vascular bundle (vb), water-storage cell (wc).



PLATE 2: D. amoenum A. Stomata, B. T.S. Leaf, C. T.S. Stem; D. amplum D. Stomata, E. T.S. Leaf, F. T.S. Stem; D. anceps G. Stomata, H. T.S. Leaf, I. T.S. Stem; D. bicameratum J. Stomata, K. T.S. Leaf, L. T.S. Stem; D. chrysanthum M. Stomata, N. T.S. Leaf, O. T.S. Stem. Stomata showing stoma (s), subsidiary cell (sc), epidermal cell (ec); T.S. Leaf showing cuticle (c), epidermis (ep), mesophyll cell (mc), xylem (x), phloem (p), fibre cap (fc), palisade layer (pl), water-storage cell (wc); T.S. Stem showing cuticle (c), epidermis (ep), ground tissue (gt), vascular bundle (vb), water-storage cell (wc).



PLATE 3: *D. crepidatum* A. Stomata , B. T.S. Leaf, C. T.S. Stem; *D. denneanum* D. Stomata , E. T.S. Leaf, F. T.S. Stem; *D. densiflorum* G. Stomata, H. T.S. Leaf, I. T.S. Stem; *D. denudans* J. Stomata, K. T.S. Leaf, L. T.S. Stem. Stomata showing stoma (s), subsidiary cell (sc), epidermal cell (ec); T.S. Leaf showing cuticle (c), epidermis (ep), mesophyll cell (mc), xylem (x), phloem (p), fibre cap (fc), water-storage cell (wc); T.S. Stem showing cuticle (c), epidermis (ep), ground tissue (gt), vascular bundle (vb), water-storage cell (wc).



PLATE 4: *D. eriiflorum* A. Stomata , B. T.S. Leaf, C. T.S. Stem; *D. fimbriatum* D. Stomata , E. T.S. Leaf, F. T.S. Stem; *D. fugax* G. Stomata, H. T.S. Leaf, I. T.S. Stem; *D. fuscescens* J. Stomata , K. T.S. Leaf, L. T.S. Stem. Stomata showing stoma (s), subsidiary cell (sc), epidermal cell (ec); T.S. Leaf showing cuticle (c), epidermis (ep), mesophyll cell (mc), xylem (x), phloem (p), fibre cap (fc), water-storage cell (wc); T.S. Stem showing cuticle (c), epidermis (ep), ground tissue (gt), vascular bundle (vb), water-storage cell (wc).



PLATE 5: *D. gibsonii* A. Stomata , B. T.S. Leaf, C. T.S. Stem; *D. heterocarpum* D. Stomata , E. T.S. Leaf, F. T.S. Stem; *D. moniliforme* G. Stomata , H. T.S. Leaf, I. T.S. Stem; *D. monticola* J Stomata , K. T.S. Leaf, L. T.S. Stem. Stomata showing stoma (s), subsidiary cell (sc), epidermal cell (ec); T.S. Leaf showing cuticle (c), epidermis (ep), mesophyll cell (mc), xylem (x), phloem (p), fibre cap (fc), water-storage cell (wc); T.S. Stem showing cuticle (c), epidermis (ep), ground tissue (gt), vascular bundle (vb), water-storage cell (wc).



PLATE 6: *D. nobile* A. Stomata, B. T.S. Leaf, C. T.S. Stem; *D. polyanthum* D. Stomata, E. T.S. Leaf, F. T.S. Stem; *D. porphyrochilum* G. Stomata, H. T.S. Leaf, I. T.S. Stem; *D. transparens* J. Stomata, K. T.S. Leaf, L. T.S. Stem. Stomata showing stoma (s), subsidiary cell (sc), epidermal cell (ec); T.S. Leaf showing cuticle (c), epidermis (ep), mesophyll cell (mc), xylem (x), phloem (p), fibre cap (fc), water-storage cell (wc); T.S. Stem showing cuticle (c), epidermis (ep), ground tissue (gt), vascular bundle (vb), water-storage cell (wc).



PLATE 7: Stegmata in Leaf surface of *Dendrobium* species (A-V). A. D. amoenum; B. D. amplum; C. D. anceps; D. D. aphyllum; E. D. bicameratum; F. D. chrysanthum; G. D. crepidatum; H. D. denneanum; I. D. denudans; J. D. eriiflorum; K. D. fimbriatum; L. D. fugax; M. D. fuscescens; N. D. gibsonii; O. D. heterocarpum; P. D. moniliforme; Q. D. monticola; R. D. moschatum; S. D. nobile; T. D. porphyrochilum; U. D. rotundatum; V. D. transparens. Leaf surface showing stegmata (s).



PLATE 8: Stegmata in Dendrobium species. T.S. Leaf (A-L) A. D. amoenum, B. D. anceps, C. D. aphyllum, D. D. bicameratum, E. D. chrysanthum, F. D. crepidatum, G. D. denudans, H. D. fugax, I. D. fuscescens, J. D. gibsonii, K. D. moschatum, L. D. nobile; T.S. Stem (M-W) M. D. amoenum, N. D. amplum, O. D. aphyllum, P. D. bicameratum, Q. D. densiflorum, R. D. formosum, S. D. fugax, T. D. longicornu, U. D. polyanthum, V. D. rotundatum, W. D. transparens. T.S. Leaf showing stegmata (s) and T.S. Stem showing stegmata (s).

Anatomical Key to the species of *Dendrobium*

1a. Leaf with thick cuticle	2
1b. Leaf with thin cuticle	10
2a. Thick cuticle with homogenous mesophyll in leaf	3
2b. Thick cuticle with heterogenous mesophyll in leaf	8
3a. Homogenous mesophyll with major vascular bundle at one leaf pole	D. anceps
3b. Homogenous mesophyll without major vascular bundle at one leaf pole	4
4a. Elongate shaped vascular bundle in stem	5
4b. Oval shaped vascular bundle in stem	D. densiflorum
5a. Stem hypodermal layers 4	D. denneanum
5b. Stem hypodermal layers < 4	6
6a. Oval shaped vascular bundle in leaf	7
6b. Flask-shaped vascular bundle in leaf	D. fugax
7a. Barrel shaped epidermal cells in stem	D. bicameratum
7b. Rectangular to polygonal shaped epidermal cells in stem	D. moniliforme
8a. Fibre cap surrounding xylem and phloem in stem	
8b. Fibre cap surrounding phloem in stem	D. rotundatum
9a. Stem vascular bundle 333.0 µm long	D. amplum
9b. Stem vascular bundle 243.09 µm long	D. fuscescens
10a. Thin leaf cuticle without hypodermis in stem	11
10b. Thin leaf cuticle with hypodermis in stem	14
11a. Fibre caps surrounding xylem and phloem present in leaf	12
11b. Fibre caps surrounding xylem and phloem absent in leaf	D. porphyrochilum
12a. Number of vascular bundles 20 in stem	D. monticola
12b. Number of vascular bundles > 20 in stem	13
13a. Stem vascular bundle 119.88 µm long	D. denudans
13b. Stem vascular bundle 199.80 µm long	D. eriiflorum
14a. Midrib vascular bundle 506.54 µm long in leaf	D. moschatum
14b. Midrib vascular bundle < 506.54 μm long in leaf	15
15a. Mesophyll layers 10 in leaf	D. crepidatum
15b. Mesophyll layers < 10 in leaf	16
16a. Phloem fibre cap 233.10 μm wide in leaf	D. gibsonii
16b. Phloem fibre cap < 233.10 μ m wide in leaf	17
17a. Xylem 16.65 µm long in stem	D. transparens
17b. Xylem > 16.65 μm long in stem	18
18a. Xylem fibre cap 86.58 μm wide in leaf	D. aphyllum
18b. Xylem fibre cap > 86.58 μ m wide in leaf	
19a. Stem vascular bundle 289.71 µm long	D. polyanthum
19b. Stem vascular bundle < 289.71 μm long	20
20a. Hypodermal layers 2 in stem	21
20b. Hypodermal layers 5 in stem	D. fimbriatum
21a. Oval shaped midrib vascular bundle in leaf	22
21b. Circular shaped midrib vascular bundle in leaf	D. longicornu
22a. Phloem fibre cap 206.46 µm wide in leaf	D. formosum
22b. Phloem fibre cap < 206.46 μ m wide in leaf	23
23a. Oval shaped vascular bundle in stem	24
23b. Elongate shaped vascular bundle in stem	D. chrysanthum
24a. Mesophyll layer 7 in leaf	25
24b. Mesophyll layer 9 in leaf	D. nobile
25a. Mesophyll cell columnar to polygonal in leaf	D. amoenum
25b. Mesophyll cell rectangular to polygonal in leaf	D. heterocarpum

Cluster analysis

The anatomical data were analyzed to find the interrelationship among the species of genus *Dendrobium*. The analysis results in one dendrogram with AU *p*-value (printed in red colour by default) and BP value (printed in green colour by default). In the dendrogram, AU *p*-value above 95% was the significant clusters.

The dendrogram of cluster analysis of species of *Dendrobium* is presented in Fig. 1. The topology showed that the species of *Dendrobium* were divided into two major clusters: cluster A with AU *p*-value of 45% and BP *p*-value of 19% and cluster B with AU *p*-value of 49% and BP *p*-value of 18% with significant values.

Cluster A was further divided into two clusters: cluster C with a single species *D. anceps* (section Aporum) and cluster D with *D. longicornu*, *D. polyanthum*, *D. porphyrochilum*, *D. denudans* and *D. transparens*. Cluster D was again divided into two clusters: cluster D1 with *D.longicornu* and *D. polyanthum* with AU *p*-value of 60% and BP *p*-value of 24% and cluster D2 with *D. porphyrochilum*, *D. denudans and D. transparens* with AU *p*-value of 68% and BP *p*-value of 24%. Cluster D2 was again divided into two clusters: cluster D2a with single species *D. porphyrochilum* (section Stachyobium) and cluster D2b with *D. denudans* and *D. transparens*.

Cluster B was further divided into two clusters: cluster E with AU *p*-value of 66% and BP *p*value of 3% and cluster F with AU *p*-value of 69% and BP *p*-value of 1%. Cluster E was again divided into two clusters: cluster E1 with *D*. *moniliforme*, *D*. *eriiflorum* and *D*. *monticola* and cluster E2 with *D*. *bicameratum*, *D*. *amoenum*, *D*. *aphyllum and D*. *heterocarpum*. Cluster E1 was again divided into two clusters: cluster E1a with a single species *D*. *moniliforme* (section Dendrobium) and cluster E1b with *D*. *eriiflorum* and *D*. *monticola*. Cluster E2 was again divided into two clusters: cluster E2a with a single species *D*. *bicameratum* (section Dendrobium) and cluster E2b with D. amoenum, D. aphyllum into two clusters: cluster E2bi with D. amoenum (section Dendrobium) and cluster E2bii with D. aphyllum and D. heterocarpum. Cluster F was divided into two clusters: cluster F1 with D. rotundatum, D. amplum and D. fuscescens (section Sarcopodium) with AU p-value of 98% and BP *p*-value of 64% with significant values and cluster F2 with D. fugax, D. formosum, D. nobile, D. densiflorum, D. moschatum, D. chrysanthum, D. gibsonii, D. crepidatum. D. denneanum and D. fimbriatum with AU p-value of 87% and BP pvalue of 1%. Cluster F1 again divided into two clusters: cluster F1a with D. rotundatum and cluster F1b with D. amplum and D. fuscescens. Cluster F2 further divided into two clusters: cluster F2a with D. fugax, D. formosum and D. nobile with AU p-value of 94% and BP p-value of 35% with significant values and cluster F2b with D. densiflorum, D. moschatum, D. chrysanthum, D. gibsonii, D. crepidatum, D. denneanum and D. fimbriatum with AU p-value of 96% and BP *p*-value of 0% with significant values. Cluster F2a again divided into two clusters: cluster F2ai with D. fugax (section Crinifera) and cluster F2aii with *D. formosum* and *D. nobile* with AU *p*-value of 92% and BP *p*-value of 54% with significant values. Cluster F2b again divided into two clusters: cluster F2bi with D. densiflorum and D. moschatum with AU p-value of 98% and BP *p*-value of 85% with significantValues and cluster F2bii with D. chrysanthum, D. gibsonii, D. crepidatum, D. denneanum and D. fimbriatum with AU p-value of 81% and BP p-value of 0% with significant values. Cluster F2bii again divided into two clusters: cluster F2biii with D. chrysanthum and D. gibsonii with AU p-value of 94% and BP pvalue of 77% with significant values and cluster F2biv with *D. crepidatum*, *D. denneanum* and *D.* fimbriatum with AU p-value of 83% and BP pvalue of 8% with significant values. Cluster F2biv again divided into two clusters: cluster F2bv with D. crepidatum and cluster F2bvi with D. denneanum and D. fimbriatum.



Figure 1. Cluster dendrogram constructed according to pvclust cluster analysis with AU/BP *p*-values in % and the bootstrapping of 10000. The AU/BP *p*-values are indicated below the branches. The AU *p*-vlues above 95% indicates the significant clusters.

Discussion

The present study revealed that the stomatal complex, stegmata, anatomy of leaves and stems of different species of *Dendrobium* showed variations in stomatal, stegmatal and anatomical characters of stem and leaves.

The stomatal complex of *Dendrobium* showed variation in types of epidermal cells, shape, size and types of stomata, stomatal frequency and stomatal index. All species of *Dendrobium* showed hypostomatic leaf surface. The leaves are found hypostamic condition in most oforchids (Avadhani *et al.*, 1982). Stomata were found only on the abaxial surface (hypostomatic). This

is because these plants are adapted to aerial habitats to minimize water loss through stomata (Stebbins and Khush, 1961). The epiderm-al cells on the abaxial surface varied from rectangular, squarish, pentagonal, hexagonal to polygonal, irregular with thick or thin walled parenchymatous cells and some species with sclerenchymatous cells. The shape of stomata varied from elliptical, circular to suborbicular. In *Dendrobium* most species had elliptical stomata and few species with circular stomata whereas in *D. amplum* and *D. rotundatum* suborbicular stomata were present. Various modifications of stomatal shape such as elliptical, circular, transversely elliptical and angular are known to exist within Orchidaceae (Rasmussen 1987). Two types of stomatal shape (Stoma I and Stoma II) were reported in *Dendrobium* (Yukawa *et al.,* 1992). In the present study, the presence of two types of stomatal shape i.e. elliptical and circular is supported by the result of Yukawa *et al.,* (1992). In addition, suborbicular stomata were found in *D. amplum* and *D. rotundatum* which are the accepted names of *Epigeneium amplum* and *Epigeneium rotundatum* according to update classification (Chase *et al.,* 2015).

Tetracytic type of stomata had been reported in many monocotyledons (Metcalfe, 1961). Mostly three types of stomata paracytic, tetracytic and anomocytric were reported in mostly in monocots (Cheadle, 1953; Stebbins and Khush, 1961). Patel (1979) distinguished five types of stomata i.e. tetracytic (a-tetra-monocyclic, b-tetra-monocyclic and c-tetra-monocyclic), twicytic, perihaplocytic, anisocytic and hexacytic and each type with four to six subsidiary cells. In the present study tetracytic type (a-tetra-monocyclic, btetra-monocyclic and c-tetra-monocyclic) was found in most species of Dendrobium whereas eupara twi-monocyclic with hexa-monocyclic types was found in one species (D. anceps). In the stomatal type the monocyclic means the guard cells of the stomata are surrounded by a single cycle of subsdiary cells (Patel, 1979). Anisocytic stomata were observed in D. amplum according to Rasmussen (1987) but it was not observed in our study.

The size of stomata showed a wide variation from 620.97 µm² to 1928.48 µm². The largest stoma was found in D. formosum and the smallest in D. fugax. The stomatal frequency ranged from 46.15 mm² to 123.07 mm². The highest stomatal frequency was found in D. longicornu and lowest in D. anceps and D. aphyllum. The reduced stomatal frequency was distinctly related with the extent of leaf succulence and more the succulence lesser the frequency of stomata (Goh et al., 1977). The stomatal index ranged from 3.75 to 12.50. The highest stomatal index was found in D. formosum and lowest in D. anceps. Similar variation in stomatal index was reported in orchid species (Saaduet al., 2009).

Stegmata was found in all species of Dendrobium except D. densiflorum, D. formosum, D. longicornu and D. polyanthum. Presence of stegmata in Dendrobiinae was supported by the result of Dressler (1993). Stegmata containing rough surfaced spherical bodies present in D. anceps was found similar to the result of Carlsward et al., (1997). Transverse section of leaf showed some variations in cuticle, shape of epidermal cells, hypodermis, mesophyll, vascular bundle and fibre caps in different species of Dendrobium.Intermittently domed cuticle was present in D. anceps. The result was found similar to the result of Morris et al., (1996). In epidermis of D. fugax, cell lumina were found occluded with Saffranin staining in adaxial and abaxial sides. A single layer of hypodermis was present in abaxial side in *D. anceps* and adaxial side in *D.* densiflorum and absent in other species. This result was found similar to the result of Morris et al (1996). Fibre bundles were present subtending the hypodermis only in D. anceps. Presence of fibre bundle in *D. anceps* is supported by the result of Morris et al., (1996). Mesophyll cells were found homogenous and heterogenous. Heterogenous mesophyll was found in D. amplum and D. rotundatum and homogenous in remai-ning species. Palisade cells were columnar in D. amplum which is supported by the result of Morris et al., (1996). Vascular bundles were arranged in a single row with large midrib bundle in most of species of *Dendrobium* except in *D. anceps* in which the vascular bundles were arranged in two rows on either side of midline with a major vascular bundle at one pole of leaf section. This result is supported by the result of Carlward et al., (1997). Dome-shaped fibre cap was present at xylem pole of D. moschatum. Transverse section of stem showed variations in cuticle, shape of epidermal cells, shape and no. of hypodermal layers, ground tissue layers, shape and no. of vascular bundles and fibre caps in different species of Dendrobium.

Cuticle is thin in *D. porphyrochilum* and thick in remaining species. Cuticle is intermittently undulating in *D. anceps*. This result was found similar to the result of Morris *et al.,* (1996). The epidermal cells vary from rectangular to square to polygonal to barrel shaped in different

species of Dendrobium. In ground tissue two kinds of water-storage cells *i.e.* pleated or unpleated were present in all Dendroboium species. Water-storage cells have been reported in Orchidaceae (Stern and Morris, 1992). Waterstorage cells with pleated or unpleated walls lack the secondary thickenings. This result is similar to the result of Morris et al., (1996). Vascular bundle shape and number varies from species to species of Dendrobium. Stegmata with rough-surfaced silica bodies were found in stems and leaves of Dendroboium species. Presence of stregmata with spherical bodies in this study is supported by the result of Kohl (1889). This result supports the hypothesis of Moller and Rasmussen (1984) in possessing stegmata with spherical silica bodies in all Dendrobiinae.

From the present study, the stomatal complex, stegmata, anatomical characters of leaves and stems were found to be taxonomically significant for delimiting the taxa within *Dendrobium* of Nepal.

Cluster analysis

Cluster analysis was carried out to find out the interrelationships among the species of Dendrobium. The cluster analysis based on anatomical with micrpmorphological characters revealed two major significant clusters: cluster A comprising 6 species with AU p-value of 45% and BP p-value of 19% belonging to sections Aporum, Formosae, Dendrobium and Stachyo-bium and cluster B comprising 20 species with AU pvalue of 49% and BP *p*-value of 18% belonging to sections Stachyobium, Dendrobium, Crinifera, Formosae and Sarcopodium. The species under cluster A were distinguished by elliptic shaped a-tetra-monocyclic, b-tetra-monocyclic to eupara-twi-monocyclic+hexa-monocyclic stomata, parenchymatous leaf epidermal cells, abaxial leaf hypodermis, homogenous mesophyll, oblong to circular midrib vascular bundle, presence or absence of fibre bundle, presence or absence of fibre cap on midrib vascular bundle of leaf, major vascular bundle at leaf pole, one to three stem hypodermal layer. In cluster B, the species were distinguished by circular to suborbicular shape with a-tetra-monocyclic, btetra-monocyclic, c-tetra-monocyclic to euparatwi-monocyclic typed stomata, parenchymatous or sclerenchymatous leaf epidermal cells, adaxial leaf hypodermis, presence of homogenous and heterogenous mesophyll, oval, flask-shaped to conical midrib vascular bundle, absence of fibre bundle and major vascular bundle at leaf pole, presence fibre cap on midrib vascular bundle of leaf, one to five stem hypodermal layer.

Cluster A was divided into two clusters: cluster C and cluster D. In cluster A, a separate cluster C was formed by D. anceps (section Aporum) as it is differed from cluster D by eupara-twimonocyclic+hexa-monocyclic stomata, thick cuticle, one layered abaxial leaf hypodermis, major vascular bundle at leaf pole and fibre bundle. In cluster D1 of cluster D, D. longicornu (section Formosae) was found close to D. polyanthum (section Dendrobium) as they had elliptic stomata, parenchymatous leaf epidermal cell, thick cuticle, circular midrib vascular bundle and elongate stem vascular bundle. In cluster D2a of cluster D, D. porphyrochilum (section Stachyobium) was found separated from cluster D2b with D. denudans (section Stachyobium) and D. transparens (section Dendrobium) as it had a-tetra-monocyclic stomata, water storage cells in mesophyll, no fibre cap on midrib bundle of leaf and stem vascular bundle. In cluster D2b, D. denudans was found close to D. transparens as both species had elliptic stomata, thin cuticle, no leaf hypodermis, homogenous mesophyll layers, stem vascular bundle number up to 60.

Cluster B was divided into two clusters: cluster E and cluster F. In cluster E1 of cluster E, *D. moniliforme* (section Dendrobium) of cluster E1a was found separated from cluster E1b in having circular shaped a-tetra-monocyclic stomata, thick cuticle and two to three polygonal stem hypodermal layers. In cluster E1b, *D. eriiflorum* (section Stachyobium) was found close to *D. monticola* (section Stachyobium) because both species had elliptic stomata, thin cuticle, no leaf hypodermis, rectangular stem epidermal cell and no stem hypodermis. In cluster E2a, *D. bicameratum* (section Dendrobium)differed from cluster E2b in having stomatal pore more than 28 µm, thick cuticle of leaf and elongate stem vascular bundle. In cluster E2bi, *D. amoenum* (section Dendrobium) differed from cluster E2bii in having b-tetra-monocyclic stomata with length up to 33 μ m, rectangular to polygonal stem epidermal cell and larger xylem in stem vascular bundle. In E2bii, *D. aphyllum* (section Dendrobium) was found close to *D. heterocarpum* (section Dendrobium) as both species had a-tetra-monocyclic stomata, thin cuticle, parenchymatous leaf epidermal cell, stegmata, no leaf hypodermis, oval midrib vascular bundle, oval stem vascular bundle and raphide bundles in stem.

In cluster F, cluster F1 with D. rotundatum, D. *amplum* and *D. fuscescens* (section Sarcopodium) form a significant cluster with AU p-value of 98% and BP p-value of 64% with significant values. D. rotundatum of cluster F1a was found separated from cluster F1b in having suborbicular c-tetra-monocyclic stomata, water storage cell and calcium oxalate crystals in mesophyll and no xylem fibre cap in stem vascular bundle. D. amplum and D. fuscescens of cluster F1bwere placed closely as both species had thick cuticle, sclerenchymatous leaf epidermal cell, stegmata, no leaf hypodermis, heterogenous mesophyll layers, flask-shaped midrib vascular bundle and two to three layers of polygonal stem hypodermis. As D. rotundatum, D. amplum and D. fuscescens were more closely related to each other, these three species may be merged into single species with further additional research. In cluster F2, D. fugax (section Crinifera) of cluster F2ai differed from cluster F2aii in having eupara-twi-monocyclic stomata, thick cuticle, more than ten homogenous mesophyll layers, flask-shaped midrib vascular bundle, barrel-shaped stem epidermal cell and elongate stem vascular bundle. In cluster F2aii, D. formosum (section Formosae) was found close to D. nobile (section Dendrobium) with AU p-value of 94% and BP p-value of 35% as both species had c-tetra-monocyclic stomata, eight to ten homogenous mesophyll layers, calcium oxalate crystals in mesophyll, oval midrib vascular bundle and two to three layers of polygonal stem hypodermis, oval stem vascular bundle. In cluster F2b, cluster F2bi formed a significant cluster with D. densiflorum and D. moschatum (section Dendrobium) with AU *p*-value of 98% and BP *p*-value of 85% with significant values. *D. densiflorum* was found close to *D. moschatum* as both species had a-tetra-monocyclic stomata, more than ten homo-genous mesophyll layers, conical midrib vascu-lar bundle, oval stem vascular bundle, stem vascular bundle no. more than 70 and raphide bundles in stem. *D. densiflorum* and *D. moscha-tum* were closely interrelated, with further research, these two species may be merged into a single species.

In cluster F2biii, D. chrysanthum (section Dendrobium) was found close to D. gibsonii (section Dendrobium) as both species had a-tetra-monocyclic stomata, thin cuticle, no leaf hypodermis, eight to ten homogenous mesophyll layers, oval midrib vascular bundle, two to three stem hypodermal layers, no xylem fibre cap in stem vascular bundle. Cluster F2biv again divided into two clusters: cluster F2bv and cluster F2bvi. Cluster F2bv with D. crepidatum (section Dendrobium) differed from cluster F2bvi with D. denneanum and D. fimbriatum in having elliptic shaped b-tetra-monocyclic stomata, parenchymatous leaf epidermal cell, phloem fibre cap layers more than 5 in midrib bundle, one stem hypodermal layer, stem vascular bundle number less than 61. In cluster F2bvi, D. denneanum was found close to D. fimbriatum as both species had circular shaped a-tetra-monocyclic stomata, sclerenchymatous leaf epidermal cell, no leaf hypodermis, barrel-shaped stem epidermal cell, four to five layers of rectangular to polygonal stem hypodermis and no xylem fibre cap in stem vascular bundle.

D. amplum, D. fuscescens and *D. rotundatum* belong to section Sarcopodium, came under genus *Epigeneium* previously but now it was merged into genus *Dendrobium* s.l. on the basis of molecular data (Schuiteman and Adams, 2014). The merging of genus *Epigeneium* into genus *Dendrobium* s.l. was seemed to be good as they formed a distinct significant cluster within genus *Dendrobium* s.l. This indicated that the present result of cluster analysis based on anatomical characters correlated with molecular data. *D. fugax* of section Crinifera came under genus *Flickingeria* in previous classification, but in update classification, the genus

Flickingeria was merged into genus *Dendrobium* s.l. based on the molecular data (Schuiteman and Adams, 2014). The inclusion of genus *Flickingeria* into genus *Dendrobium* s.l. was seemed correct as *D. fugax* showed close relationship with other species of *Dendrobium*. This cluster analysis result revealed that the analysis result based on anatomical characters with micromorphological characters correlate with molecular data.

In the present study, according to result of cluster analysis based on anatomical characters with micro-morphological characters, *Dendrobium* species of Nepal can be divided into new 6 sections with closely interrelated species. Sections are as follows:

Section I. includes species with eupara-twimonocyclic+hexa-monocyclic stomata, one layered abaxial leaf hypodermis, major vascular bundle at leaf pole and fibre bundle. ------*D. anceps*

Section II. includes species with elliptic stomata and small stem vascular bundle less than 100 μm ----- *D. longicornu*, *D. polyanthum*, *D.porphyrochilum*, *D.denudans* and *D. transparens*

Section III. includes species with oval midrib vascular bundle and no xylem fibre cap in stem vascular bundle -----D. moniliforme, D. eriiflorum, D. monticola, D. bicameratum, D. amoenum, D. aphyllum and D. heterocarpum

Section IV. includes species with circular or suborbicular stomata, sclerenchymatous leaf epidermal cell, heterogenous mesophyll layer and rectangular to square stem epidermal cell -- *D. rotundatum*, *D. amplum* and *D. fuscescens*

Section V. includes species with stomatal index more than 10%, xylem fibre cap in midrib vascular bundle more than 150 µm in width and polygonal stem hypodermal cell -----------*D. fugax, D. formosum* and *D. nobile*

Section VI. includes species with a-tetramonocyclic stomata ------D. densiflorum, D. moschatum, D. chrysanthum, D. gibsonii,D. crepidatum, D. denneanum and D. fimbriatum but b-tetramonocyclic stomata - D. crepidatum, no leaf hypodermis but with one layered adaxial leaf hypodermis - *D. densiflorum*, large stem vascular bundle width more than 130 μ m but less than 100 μ m - *D. crepidatum* and no stegmata but present - *D. densiflorum*.

Conclusion

The present study was based on anatomical characters of leaves and stems of *Dendrobium* species of Nepal Himalaya. Based on characters such as stomatal complex, stegmata and anatomical characters of leaves and stems, an anatomical key was prepared to delimit the taxa within genus *Dendrobium*.

Based on anatomical with micromorphological characters, the cluster analysis was carried out in species of Dendrobium to find out the interrelationships among the species of Dendrobium. With the result of cluster analysis, Dendrobium of Nepal was divided into 6 sections: section I with D. anceps, section II with D. longicornu, D. polyanthum, D. porphyrochilum, D. denudans, D. transparens, section III with D. moniliforme, D. eriiflorum, D. monticola, D. bicameratum, D. amoenum, D. aphyllum, D. heterocarpum, section IV with D. rotundatum, D. amplum, D. fuscescens, section V with D. fugax, D. formosum, D. nobile and section VI with D. densiflorum, D. moschatum, D. chrysanthum, D. gibsonii, D. crepidatum, D. denneanum and D. fimbriatum.

The present study represents the first attempt to bring the cluster analysis of *Dendrobium* of Nepal based on anatomical with micromorphological characters. The proposed classification based on the cluster analysis is slightly similar to classification of Schuiteman and Adams (2014).

Due to unavailability of the modern equipment like Scanning Electron Microscope (SEM), Transmission Electron Microscope (TEM) and DNA sequences, a detailed study of the species could not be carried out. However, the result obtained from the present study will provide the working hypothesis for further study. Therefore, it is recommended that additional researches should be needed to fully resolve the relationships between several species of *Dendrobium*.

Acknowledgements

Authors are thankful to Suraj Balami for his help in processing cluster analysis, National Herbarium and Plant Laboratories (KATH) and Tribhuvan University Central Herbarium (TUCH) for their cooperation and allowing to studying the herbarium specimens housed over there. My sincere thanks go to Patan Multiple Campus and Amrit Campus for providing the laboratories facilities.

References

- 1. Arditti, J. "Fundamental of orchid biology". John Wiley and Sons, New York, USA. (1992). Print.
- Avadhani, P. N., C. J. Goh, A. N. Rao and J. Arditti. "Carbon Fixation in Orchids". In: Orchid Biology Reviews & Perspectives II. (ed. J. Arditti) (1982): 173-93. Online
- Bajracharya, D. M. "The genus *Eria* Lindley (Orchidaceae) in the Himalayas - A taxonomic revision". Ph.D Thesis, Central Department of Botany, Tribhuvan University, Kirtipur, Nepal. (2003). Print.
- Carlsward, B. S., W. L. Stern, W. S. Judd and T. W. Lucansky. "Comparative leaf anatomy and systematics in Dendrobium sections Aporum and Rhizobium (Orchidaceae)". *International Journal of Plant Sciences* 158.3 (1997): 332–342. Online.
- 5. Carpenter, K. J. "Stomatal architecture and evolution in basal angiosperms". *American Journal of Botany* 92.10 (2005): 1595-1615. Online.
- Chase, M. W., K. M. Cameron, J. V. Freudenstein, A. M. Pridgeon, G. Salazar, C. van denBerg and A. Schuiteman. "An updated classification of Orchidaceae". *Botanical Journal* of the Linnean Society 177 (2015): 151–174. Online.
- Cheadle, V. I. "Independent origin of vessels in the monocotyledons & dicotyledons". *Phytomorphology* (1953): 23-44. Online.
- 8. Cutler, D. F. "Applied plant anatomy". Longman group limited, UK. (1978). Print.
- Dangol, G. "Study of anatomical structure of leaf and root of subtribe Coelogyninae Bentham (Orchidaceae) in Nepal". M. Sc. thesis submitted to T.U., Nepal. (2006). Print.

- 10. Dressler, R. L. "Phylogeny and Classification of the Orchid Family". Cambridge University Press, Cambridge MA, USA. (1993).
- 11. Goh, C. J., P. N. Avadhani, C. S. Loh, C. Manegraff and C. Arditti. "Diurnal stomatal and acidic rhythms in orchid leaves". *New Phytologist* 78 (1977): 365-72. Online.
- 12. Holttum, R. E., F. G. Brieger and P. J. Cribb. "A proposal for the re-typification of *Dendrobium* Sw. nom. cons". *Taxon* 28.4 (1979): 409. Online.
- Johansen, D. A. "Plant Microtechnique". Tata McGraw-Hill Book Company, India. (1940): 523. Print.
- 14. Kaushik' P. "Ecological and anatomical marvels of the Himalayan orchids". Today and Tomorrow's Printers and Publishers, New Delhi, India. (1983). Print.
- 15. Khasim, S. M. and P. R. M. Rao. "Anatomy in relation to taxonomy in some members of *Epidendroideae* (Orchidaceae)". *Phytomorphology* 40.3-4 (1990): 243-250. Online.
- 16. Kohl, F. G. "Anatomisch-Physiologische Untersuc-hung der Kalksalze und Kiselsaure in der Pfla-nze". Marburg, Germany. (1889). Print.
- 17. Link, H. F. "Bemerkungen iber den Bau der Orchideen, besonders der Vandeen". *Botanische Zeitung* 7 (1849): 745-750. Online.
- 18. Liu, W. and X. Zhu. "Leaf epidermal characters and taxonomic revision of Schizophragma and Pileostegia (Hydrangeaceae)". *Botanical Journal of the Linnean Society* 165 (2011): 285–314. Online
- Metcalfe, C. R. "The anatomical approach to systematic: General introduction with special reference to recent work on monocotyledons". *In: Recent Advances in Botany*. University of Toronto Press, Canada. (1961): 146-150. Online.
- Khasim, S. M. and P. R. Mohana Rao. "Anatomy in relation to taxonomy in some members of Epidendroideae (Orchidaceae)". *Phytomorphology* 40 (1990): 243–250. Online.
- 21. Moller, J. D., H. Rasmussen. "Stegmata in Orchidales: character state distribution and polaity". *Botanical Journal of the Linnean Society* 89 (1984): 53-76. Online.
- 22. Morris, M. W., W. L. Stern and W. S. Judd. "Vegetative anatomy and systematics of subtribe *Dendrobiinae* (Orchidaceae)". *Botanical Journal of the Linnean Society* 120 (1996): 89-144. Online.

- 23. Patel, J. D. "New morphological classification of stomatal complexes". *Phytomorphology* 29 (1979) : 218-229. Online.
- 24. Rajbhandari, K. R., S. Bhattarai and R. Joshi. "Orchids Diversity in Nepal and Their Conservation". In: Proceedings of 8thInternational Conference on BIO-REFOR, Nepal. (1999). Print.
- Rajbhandari, K. R. "Orchids of Nepal: Status, Threat and conservation". In: Proceeding of National workshop on NTFP/ Maps sector action plan development: Orchid. Dept. of Plant Resources and Tribhuvan University, Nepal. 1 (2014): 1–40. Print.
- Rasmussen, H. "Orchid stomata-structure, differentiation, function and phylogeny". *In*: Arditti, J. (ed.) *Orchid biology-reviews and perspectives*. Cornell University Press, Ithaca, New York, USA. 4 (1987): 105-138. Print.
- Saadu, R. O., A. A. Abdulrahaman and F. A. Oladele. "Stomatal complex types and transpiration rates in some tropical tubers species". *African Journal of Plant Science* 3.5 (2009): 107-112. Online.
- 28. Salisbury, E. J. "On the causes and ecological significance of stomatal frequency with reference to the woodland flora". *Philosophical Transactions of the Royal Society* 216 B (1928): 1-65. Online.
- Schuiteman, A. "Dendrobium". In: A. M. Pridgeon, P. J. Cribb and M.W. Chase (eds.) Genera Orchidacearum vol: 6 Epidendroideae (Part Three). Oxford University Press, Oxford, UK. (2014): 51-73. Print.
- Schuiteman, A and P. B. Adams. "Tribe Dendrobieae". In: A. M. Pridgeon, P. J. Cribb, M. W. Chase and F. N. Rasmussen (eds.). Genera Orchidacearum vol. 6 Epidendroideae (Part

three). Oxford University Press, Oxford, U. K. (2014): 3-4, 51-73, 89-100. Print.

- 31. Shakya, L. R. "Revision of the genus *Oberonia* Lindl. (Orchidaceae) in the Himalayas". Ph.D. Thesis, Central Department of Botany, Tribhuvan University, Kirtipur, Nepal. (1999). Print.
- 32. Singh, V. and H. Singh. "Organisation of stomatal complex in some Orchidaceae". *Current Science* 43 (1974): 490-491. Online.
- 33. Stebbins, G. L., G. S. Khush. "Variation in the organization of the stomatal complex in the leaf epidermis of monocotyledons and its bearing on the phylogeny". *American Journal of Botany* 48.1. (1961): 51-59. Online.
- 34. Stern, W. L. and M. W. Morris. "Vegetative anatomy of *Stanhopea* (Orchidaceae) with special reference to pseudobulb water-storage cells". *Lindleyana* 7 (1992): 34-53. Online.
- 35. Suzuki, R. and H. Shimodaira. "Pvclust: hierarchical clustering with p-values via multiscale bootstr-ap resampling". http:// CRAN.R-project.org/package=pvclust,R package ver.1.2-2. (2011). Online.
- Swartz, O. "Dendrobium". Nova Acta Regiae Soci-etatis Scientiarum Upsaliensis 6.2 (1799): 82-85. Online.
- 37. Vij, S. P., P. S. Kaushal and P. Kaur. "Observations on leaf epidermal features in some Indian orchids: taxonomic and ecological implications". The *Journal of the Orchid Society of India* 5 (1991): 43-53. Online.
- Yukawa, T., T. Ando, K. Karasawa, K. Hashimoto. "Existence of two stomatal shapes in the genus *Dendrobium* (Orchidaceae) and its systematic significance". *American Journal of Botany* 79.8 (1992): 946-952. Online.

Cite this article as:

Baba Maiya Pradhan and Devendra M. Bajracharya. Anatomical Study of *Dendrobium* (Orchidaceae) of Nepal. *Annals of Plant Sciences*. 9.7 (2020) pp. 3916-3948.

bttp://dx.doi.org/10.21746/aps.2020.9.7.1

Subject Editor : Mitta Mahendra Nath, Sri Venkateswara University, Tirupati.

Source of support: National Herbarium and Plant Laboratories (KATH) and Tribhuvan University Central Herbarium (TUCH) Conflict of interest: Nil.