



2D QSAR STUDY ON SERIES OF TACRINE AND RELATED COMPOUNDS WITH ACETYLCHOLINESTERASE INHIBITORY ACTIVITY

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Abstract

Introduction: Acetylcholine esterase inhibitors are the drugs that provide symptomatic treatment for Alzheimer's disease. The AChE inhibitors, which interact with both, peripheral and active site of the enzyme, can also inhibit the formation of β A4-amyloid protein (β AP) in addition to AChE inhibition. In order to gain a deeper insight of drug receptor interaction QSAR study has been performed.

Material and Methods: Compounds reported in the literature has been taken up for the study. The physicochemical parameters were calculated and it was then correlated with the given acetylcholinesterase activity using Systat 7 statistical software.

Results: A series of 77 structurally diverse group AChE inhibitors reported in the literature have been investigated and a comprehensive QSAR study has been performed. A meaningful QSAR equation has been derived which explains the hydrophobic and molar refractivity as a key physicochemical parameters contributing towards activity.

Conclusion: The QSAR model developed in this study can provide a better insight of the physicochemical parameters and can be useful in designing novel acetylcholinestrase inhibitors.

Keywords: Alzheimers, Acetylcholinestrase inhibition, 2D QSAR, regression analysis.

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INTRODUCTION

Alzheimer's disease is a devastating degenerative disorder of the central nervous system that results in gradual deterioration of cognitive function and severe alteration of personality^(1,2) Degeneration of neurons in the nucleus basalis Meynert, the origin of the major cholinergic projections to the neocortex,⁽³⁻⁶⁾ occurs early in the course of the disease, and is correlated with the cognitive decline. The biochemical deficit also extends to other neurochemicals systems affecting the level of monamine transmitter,⁷ but cholinergic neurotransmission is the one that is greatly affected. Using *in-vivo* rodent models, several cholinergic enhancement strategies including neuropharmacological approaches (acetylcholinesterase inhibitors), neurotrophic factor administration (nerve growth factor),⁸ and transplantation of cholinergic-enriched fetal grafts have been developed and were proven to be effective in alleviating lesion-induced cognitive

deficits. On the basis of preclinical and clinical studies cholinesterase inhibitors are the only approved drugs for the treatment of cognitive deficit associated with Alzheimers disease.⁹ The AChE inhibitors restore deficient cholinergic neurotransmission. by increasing the available acetylcholine within the active synapse Increasing the level of acetylcholine has been regarded as one of the most promising methods for the palliative treatment of AD^(10,11) Indeed, the three drugs (Donepezil, Rivastigmine, Galanthamine currently approved for the treatment of Alzheimer's disease are cholinomimetics with the pharmacological profile of acetylcholinesterase inhibitors^(12,13) These drugs are prescribed to treat symptoms related to memory loss, thinking, language judgments and other thought processes. Beside these three acholinesterase inhibitors, another drug Memantine¹⁴ has been approved to treat moderate to severe Alzheimer's. It is the first novel class of Alzheimer's disease medications acting on

the glutamatergic system by blocking NMDA-type glutamate receptors.

Extensive efforts by different researchers have led to discovery of a number of potent AChE inhibitors with structural diversity, such as xanthostigmine, physostigmine, phenserine, huperzine-A, bis-tacrine, bis-huperzin-B⁽¹⁵⁻²⁰⁾. Studies have shown that the AChE inhibitors, which interact with both, peripheral and active site of the enzyme⁽²¹⁻²⁴⁾ can also inhibit the formation of β A4-amyloid protein (β AP) in addition to AChE inhibition. This could be achieved by blocking the active site of acetylcholinesterase enzyme. Tacrine was launched as the first drug for the treatment of Alzheimers related symptoms. However it is no longer used because of its hepatotoxicity. In order to improve its efficacy, its several analogs have been synthesized and evaluated for their AChE inhibitory activity. The comparative QSAR studies on most of these compounds have been reported by Hanch and Maurizino²⁵. However in these studies they could not draw any conclusion about the effect of steric and hydrophobic parameters on activity because of high colinearity between these parameters. Therefore they suggested about the need of carefully designed analogs for filling the gap in the parameter space and for getting the sound basis for building more informative QSAR. Later Maurizino et al²⁶ synthesized tetrahydroacridine analogs with non collinear substituents in terms of steric and hydrophobic parameters. However the best 2D and 3D QSAR models developed for these and related 11H-Indeno [1, 2-b] quinolin-10ylamine analogs, did not include the N-9-substituted compounds even though they had high activity. The reason given for their exclusion was that the conformational flexibility of both benzyl and heptyl groups at N-9 position of acridine had made the choice of proper physicochemical parameter questionable. Moreover in these models as well as in the docking studies neither N-9-substituted nor 1-hydroxy (velnacrine) analogs were considered. Hence it appeared of interest to relook into these studies for the development of comprehensive 2D models taking into consideration the issues of colinearity between the steric and hydrophobic parameter and the inclusion of N-9 substituted and 1-hydroxy analogs in the study.

Structure of the inhibitors Fig. 1

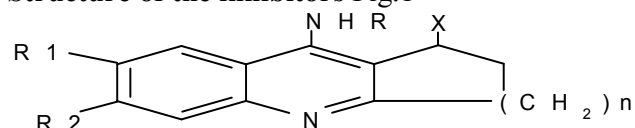


Fig. 1

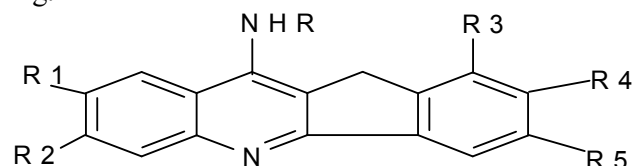


Fig. 2

This study was undertaken to develop a correlation between acetylcholinesterase activity of tacrine analogs with different physicochemical parameters.

MATERIAL AND METHODS

This study was conducted at Central Drug Research Institute for a period of six months. The reported data of 77 compounds from three different sources⁽³⁵⁻³⁷⁾ was considered for QSAR analysis. The AChE inhibitory (IC_{50}) activities of these compounds were from two different labs; however compounds from both the labs were tested using Ellman's protocol. Since the activity of the standard drug tacrine was different in the dataset reported from two different groups being $0.319 \mu M$ in the dataset reported by Shustke et al and $0.25 \mu M$ in the other two datasets. Hence the activities of the compounds reported in the latter two data sets were normalized by multiplying them by the factor of 1.27 ($tac1/tac2m \ 0.319/0.25=1.27$) (Table 1,2)

2D QSAR Development

The preliminary 2D QSAR analysis was carried out using the AChE inhibitory activity of all the 77 compounds as dependent and the physicochemical parameters such as hydrophobicity (π), steric (E_s, MR) and electronic (R, F, σ) and the indicator variables as the independent parameters. The values of different physicochemical parameters were computed from the literature values⁽³⁸⁾ and the two indicator parameters I_1 and I_2 were used for the presence of hydroxyl group ($I_1=1$) and the tetrahydroacridine nucleus ($I_2=1$) respectively. The multiparameter analysis was carried using Systat software 7.0

RESULTS

2D QSAR

The multiparameter regression analysis (MLR) using different permutation and combinations of the independent parameters viz hydrophobicity (π), steric (E_s , MR) and electronic (R , F , σ) and the indicators variables led to the development of the 2D QSAR model (eq1), where the figures in parentheses describe the standard error of the regression coefficient, n represents the number of data points, r is the correlation coefficient, s is the standard error from the regression and F is the measure of statistical significance of the regression model.

$$-\log IC_{50} = 0.798(\pm 0.220) \pi R_2 - 0.179(\pm 0.042) MR_1 - 0.019(\pm 0.005) MRR - 1.284(\pm 0.171) I_1 + 1.251(\pm 0.215) I_2 - 0.282$$

$$n=77, \quad r=0.784, \quad s=0.584, \quad F=22.760$$

eq1

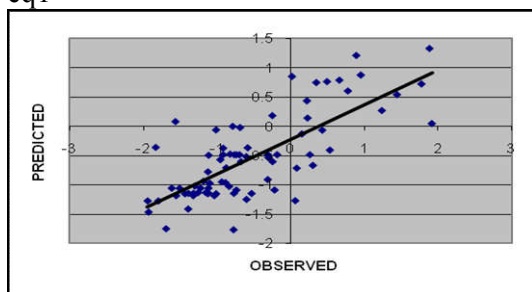


Fig 3 Graph showing linear relationship between the observed and predicted values. This model with moderate correlation coefficient value (0.784), low standard error (0.584), high statistical significance ($F_{5, 71} \alpha 0.001 = 5.08$; $F_{5, 71} = 22.760$) and with >99.9% values for regression coefficients well describes the variation in the observed acetylcholinesterase inhibitory activity of all the 77 compounds of the data set. as shown by comparison of observed and calculated acetylcholinesterase inhibitory activity of these molecules (table 3, fig.3). It is interesting to note that there is no intercorrelation between the independent parameters ($r < 0.5$) (Table 4).

Table 4: Correlation matrix (r values) of physicochemical parameters

	- LOGIC ₅₀	πR_2	MR_1	MRR	I_1	I_2
-LOGIC ₅₀	1.000					
πR_2	0.389	1.000				
MR_1	0.004	0.059	1.000			
MRR	-0.323	0.029	-0.243	1.000		
I_1	-0.477	0.047	-0.318	0.469	1.000	
I_2	0.102	0.229	0.135	0.453	0.463	1

DISCUSSION

The issues of co-linearity between the steric and hydrophobic parameter and the inclusion of N-9 substituted and 1-hydroxy analogs of tacrine which was not reported in the earlier studies was resolved.⁽²⁵⁻²⁸⁾ It is evident from the equation that hydrophobicity and steric effects play an important role in the variation of activity. Thus the hydrophobicity of the substituents (R_2) at 6th position for tacrine and velnacrine and 7th position of 11H-Indeno[1,2-b] quinolin-10ylamine analogs contributes positively while the increase in bulk at 7th position (R_1) for tacrine and velnacrine and 8th position of 11H-Indeno[1,2-b] quinolin-10ylamine contributes negatively for the activity. This indicates that for molecule to possess acetylcholinesterase inhibitory activity^(30,31) the molecule must be hydrophobic in nature. The result also indicates that bulky substituents at the specified position will be detrimental for the activity. Further the presence of hydroxyl group in the nucleus in the prototypes ($I_1=1$) and tetrahydroacridine ($I_2=1$) contributes negatively and positively respectively thus indicating that the tetrahydroacridine nucleus devoid of the hydroxyl group best suits for activity.

CONCLUSION

The comprehensive QSAR performed in this study has given a better insight about the structural modification that can be incorporated, in order to produce novel molecule with acetylcholinesterase activity. Further exploration with different scaffold will enhance better understanding of contributing physicochemical parameters towards acetylcholinesterase activity.

Additional files

Additional File – 1: Table 1 – Data Set 1 Tacrine Related Molecules

Additional File – 2: Table 2 – Data Set 2 Tacrine Related Molecules

Additional File – 3: Table 3 – Physicochemical 2D Parameters

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REFERENCES

- Drevets WC, Rubin EH. Psychotic symptoms and the longitudinal course of senile dementia of the Alzheimer type. *Biol. Psychiatry.* 1989;25:39-48.
- Bartus RT, Dean RL 3rd, Beer B, Lippa AS. The cholinergic hypothesis of geriatric memory dysfunction. *Science.* 1982;217: 408-14

3. Mesulam MM, Mufson EJ, Levey AI, Wainer BH.J. Cholinergic innervation of cortex by the basal forebrain: cytochemistry and cortical connections of the septal area, diagonal band nuclei, nucleus basalis (substantia innominata), and hypothalamus in the rhesus monkey. *Comp. Neurol.*1983 ;214:170- 97
4. Mesulam MM, Geula C. Nucleus basalis (Ch4) and cortical cholinergic innervation in the human brain: observations based on the distribution of acetylcholinesterase and choline acetyltransferase. *J Comp Neurol.* 1988;275:216–240.
5. Mesulam MM, Hersh LB, Mash DC, Geula C. Differential cholinergic innervation within functional subdivisions of the human cerebral cortex: a choline acetyltransferase study. *J Comp Neurol.*1992a;318:316–328
6. Hedreen JC, Struble RG, Whitehouse PJ, Price DL. Topography of the magnocellular basal forebrain system in human brain. *J Neuropathol Exp Neurol* 1984;43:1-21.
7. Rossor M, Iversen LL. Non-cholinergic neurotransmitter abnormalities in Alzheimer's disease. *Br Med Bull.* 1986 Jan;42(1):70–74.
8. Yong C, Youhua S, Dexing FU. Drug therapy for Alzheimers Disease The Chinese Journal of Clinical Pharmacology.1999;4:295-298
9. Siddiqui, MF, Levey, AI. Cholinergic therapies in Alzheimer's disease. *Drugs Fut.* 1999;24: 417-444
10. Enz A, Amstutz R, Boddeke H, Gmelin G, Malanowski J. Brain selective inhibition of acetylcholinesterase: a novel approach to therapy for Alzheimer's disease. *Prog Brain Res.* 1993; 98:431-8.
11. Millard CB, Broomfield CA. Anticholinesterases: medical applications of neurochemical principles. *J. Neurochem.* 1995; 64:1909-1918.
12. Lahiri DK, Rogers JT, Greig N H, Sambamurti K. Rationale for the development of cholinesterase inhibitors as anti-Alzheimer agents. *Curr. Pharm Des.*2004;10: 3111–19.
13. Pepeu G, Giovannini M G. 2009. Cholinesterase inhibitors and beyond. *Curr. Alzheimer Res.*2009; 6: 86–96.
14. Reisberg B, Doody R, Stöffler A, Schmitt F, Ferris S, Möbius HJ, Memantine Study Group, Memantine in moderate-to-severe Alzheimer's disease. *N Engl J Med.* 2003 Apr 3;348(14):1333-1341.
15. Rampa A, Bartolini M, Bisi A, Belluti F, Gobbi S, Andrisano V, et al. The First Dual ChE/FAAH Inhibitors: New Perspectives for Alzheimer's Disease? *Med Chem Lett.* 2012 Jan 21; 3(3):182-6.
16. Möller H-J, Hampel H, Hegerl U, Schmitt W, Walter K. Double-blind, randomized, placebo-controlled clinical trial on the efficacy and tolerability of a physostigmine patch in patients with senile dementia of the Alzheimer type. *Pharmacopsychiatry.* 1999 May; 32(3):99-106.
17. Winblad B, Giacobini E, Frölich L, Friedhoff LT, Bruinsma G, Becker RE, et al. Phenserine efficacy in Alzheimer's disease. *J Alzheimers Dis.* 2010;22(4):1201-8.
18. Rafii MS, Walsh S, Little JT, Behan K, Reynolds B, Ward C, et al. Alzheimer's Disease Cooperative Study. A phase II trial of huperzine A in mild to moderate Alzheimer disease. *Neurology.* 2011; 76(16):1389-94.
19. Pang YP, Quiram P, Jelacic T, Hong F, Brimijoin S. Highly potent, selective, and low cost bis-tetrahydroaminacrine inhibitors of acetylcholinesterase. Steps toward novel drugs for treating Alzheimer's disease. *J Biol Chem.* 1996 Sep 27;271(39):23646-9.
20. Feng S., Wang Z., He X., Zheng S., Xia Y., Jiang H., et al. Bis-huperzine B: highly potent and selective acetylcholinesterase inhibitors. *J. Med. Chem.* 2005;48:655–657.
21. Giacobini E, Mori F, Lai CC. The effect of cholinesterase inhibitors on the secretion of APPS from rat brain cortex. *Ann N Y AcadSci.* 1996; 777: 393-98.
22. Taylor P, Lappi S. Interaction of fluorescence probes with acetylcholinesterase. The site and specificity of propidium binding. *Biochemistry.* 1975 May 6;14(9):1989–1997
23. Muñoz-Ruiz P, Rubio L, García-Palomero E, Dorronsoro I, del Monte-Millán M, Valenzuela R, et al. Design, synthesis, and biological evaluation of dual binding site acetylcholinesterase inhibitors: new disease-modifying agents for Alzheimer's disease. *J. Med. Chem.* 2005; 48:7223-7233
24. Shaik JB, Palaka BK, Penumala M, Eadlapalli S, Darla Mark M, Ampasala DR, et al. Synthesis, Biological Evaluation, and Molecular Docking of 8-imino-2-oxo-2H,8H-pyrano[2,3-f]chromene Analogs: New Dual AChE Inhibitors as Potential Drugs for the Treatment of Alzheimer's Disease. *Chem Biol Drug Des.* 2016 Jul;88(1):43-53.
25. Quinn DM. Acetylcholinesterase: enzyme structure, reaction dynamics, and virtual transition states. *Chem. Rev.* 1987; 87 (5): 955–979
26. Massoulie J, Pezzementi L, Bon S, Krejci E, Vallette FM. Molecular and cellular biology of cholinesterases. *Prog Neurobiol.* 1993;41:31-91.
27. Harel, M., Quinn, D.M., Nair, H.K., Silman, I. & Sussman, J.L. The structure of a transition state analog complex reveals the molecular origins of the catalytic power and substrate specificity of acetylcholinesterase. *J. Am. Chem. Soc.* 1996;118: 2340-2346.
28. Rosenberry TL. Acetylcholinesterase. *Adv. Enzymol. Relat. Areas. Mol. Biol.* 1975; 43: 103-218
29. Rosenberry T L. Bernhard, S.A. Studies of catalysis by acetylcholinesterase. Synergistic effects of inhibitors during the hydrolysis of acetic acid esters. *Biochemistry.* 1972; 11:4308-4321
30. Radic Z, Duran R, Vellom DC, Li Y, Cervenansky C, Taylor P. Site of fasciculin interaction with acetylcholinesterase. *J. Biol. Chem.* 1994; 269: 11233–11239
31. Shafferman A, Velan B, Ordentlich A. Kronman C, Grosfeld H, Leitner M, et al. Substrate inhibition of acetylcholinesterase: residues affecting signal transduction from the surface to the catalytic center. *EMBO J.* 1992 Oct; 11(10): 3561–3568.
32. Barak D, Kronman C, Ordentlich A, Ariel N, Bromberg A, Marcus D, et al. Acetylcholinesterase peripheral anionic site degeneracy conferred by amino acid arrays sharing a common core. *J Biol Chem.* 1994 ;269(9):6296-305.
33. Radić Z, Quinn DM, Vellom DC, Camp S, Taylor P. Allosteric control of acetylcholinesterase catalysis by fasciculin. *J Biol Chem.* 1995;270(35):20391-9.
34. Recanatini M, Cavalli A, Hansch C. A comparative QSAR analysis of acetylcholinesterase inhibitors currently studied for the treatment of Alzheimer's disease. *Chem Biol Interact.* 1997; 105(3):199-228.
35. Recanatini M, Cavalli A, Belluti F, Piazzi L, Rampa A, Bisi A, et al. SAR of 9-amino-1, 2, 3, 4-tetrahydroacridine-based acetylcholinesterase inhibitors: synthesis, enzyme inhibitory activity, QSAR, and structure-based CoMFA of tacrine analogues. *J. Med. Chem.*2000;43 (10): 2007-2018
36. Shutske G M, Pierrat FA, Kapples K J, Cornfeldt ML, Szewczak MR, Huger FP, et al. 9-Amino-1,2,3,4-tetrahydroacridin-1-ols. Synthesis and evaluation as potential Alzheimer's disease therapeutics. *J. Med. Chem.* 1989; 32: 1805–1813
37. Rampa A, Bisi A, Belluti F, Gobbi S, Valenti P, Andrisano V al. Acetylcholinesterase inhibitors for potential use in Alzheimer's disease: molecular modeling, synthesis and kinetic evaluation of 11H-indeno-[1, 2-b]-quinolin-10-ylamine derivatives. *Bioorganic & medicinal chemistry.*2000; 8 (3):497-506
38. Hansch C, Leo A. Substituent Constants for Correlation Analysis in Chemistry and Biology, John Wiley & Sons, New York, 1979.

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Table 1

Compd no	R	R1	R2	n	X	IC ₅₀ (a)	IC ₅₀ (b)	-log
1	H	H	H	2	OH	4.00	4.00	-0.602
2	H	Cl	H	2	OH	24.70	24.70	-1.393
3	H	H	Cl	2	OH	0.0117	0.0117	1.921
4	H	H	CH ₃ O	2	OH	2.00	2.00	-0.301
5	H	H	CF ₃	2	OH	1.77	1.77	-0.248
6*	H	H	F	2	OH	0.292	0.292	0.535
7*	H	H	H	1	OH	5.97	5.97	-0.776
8*	CH ₃	H	H	2	OH	1.76	1.76	-0.245
9*	N-C ₂ H ₅	H	H	2	OH	13.3	13.3	-1.124
10	(CH ₃) ₂ N(CH ₃) ₂	H	H	2	OH	7.68	7.68	-0.885
11	(CH ₃) ₂ C ₆ H ₄	H	H	2	OH	3.39	3.39	-0.53
12*	(CH ₃) ₂ OC ₆ H ₄	H	H	2	OH	3.98	3.98	-0.599
13	(CH ₃) ₂ CH(C ₆ H ₄) ₂	H	H	2	OH	49.3	49.3	-1.693
14*	(CH ₃) ₂ CH(4FC ₆ H ₄) ₂	H	H	2	OH	87.8	87.8	-1.943
15	(CH ₃) ₂ CH(3-	H	H	2	OH	63.0	63.0	-1.799
16	CH ₂ (C ₆ H ₅) ₂	H	H	2	OH	32.2	32.2	-1.508
17	CH ₂ (2-ClC ₆ H ₄) ₂	H	H	2	OH	10.3	10.3	-1.013
18*	CH ₂ (3-ClC ₆ H ₄) ₂	H	H	2	OH	13.3	13.3	-1.124
19	CH ₂ (4-ClC ₆ H ₄) ₂	H	H	2	OH	27.2	27.2	-1.435
20	CH ₂ (2-FC ₆ H ₄) ₂	H	H	2	OH	41.3	41.3	-1.616
21	CH ₂ (3-FC ₆ H ₄) ₂	H	H	2	OH	13.1	13.1	-1.117
22	CH ₂ (4-FC ₆ H ₄) ₂	H	H	2	OH	16.7	16.7	-1.223
23	CH ₂ (2-CH ₃ OC ₆ H ₄) ₂	H	H	2	OH	36.0	36.0	-1.556
24	CH ₂ (3-CH ₃ OC ₆ H ₄) ₂	H	H	2	OH	21.1	21.1	-1.324
25*	CH ₂ (4-CH ₃ OC ₆ H ₄) ₂	H	H	2	OH	10.9	10.9	-1.037
26*	CH ₂ (2-CH ₃ C ₆ H ₄) ₂	H	H	2	OH	5.89	5.89	-0.77
27*	CH ₂ (3-CH ₃ C ₆ H ₄) ₂	H	H	2	OH	24.5	24.5	-1.389
28	CH ₂ (4-CH ₃ C ₆ H ₄) ₂	H	H	2	OH	21	21	-1.322
29	CH ₂ (2-CF ₃ C ₆ H ₄) ₂	H	H	2	OH	13.3	13.3	-1.124
30	CH ₂ (2-CF ₃ C ₆ H ₄) ₂	H	H	2	OH	14.2	14.2	-1.152
31	CH ₂ (4-CF ₃ C ₆ H ₄) ₂	H	H	2	OH	18.1	18.1	-1.258
32	CH ₂ C ₆ F ₅	H	H	2	OH	12.6	12.6	-1.1
33	CH ₂ (2-C ₆ H ₄ S)	H	H	2	OH	15.3	15.3	-1.185
34	CH ₂ C ₆ H ₄	H	Cl	2	OH	1.30	1.30	-1.114
35	CH ₂ (4-FC ₆ H ₄) ₂	H	Cl	2	OH	0.823	0.823	0.084
36	CH ₂ C ₆ H ₄	H	F	2	OH	8.63	8.63	-0.936
37*	CH ₂ (2-CFC ₆ H ₄) ₂	H	F	2	OH	20.1	20.1	-1.303
38	CH ₂ C ₆ H ₅	H	CF ₃	2	OH	68.2	68.2	-1.834
39	H	CH ₃	H	2	H	8.1	10.33	-1.014
40	H	H	CH ₃	2	H	0.1	0.127	0.894
41	H	Cl	H	2	H	0.55	0.7018	0.154
42	H	H	Cl	2	H	0.0099	0.126	1.889
43	H	NO ₂	H	2	H	3	3.828	-0.583
44*	H	H	NO ₂	2	H	0.028	0.0357	1.447
45	H	H	OCH ₃	2	H	0.35	0.4466	0.35
46*	H	NH ₂	H	2	H	3.8	4.8488	-0.686
47	H	H	F	2	H	0.087	0.1110	0.955
48	H	Cl	Cl	2	H	0.47	0.5997	0.222
49	H	OCH ₃	OCH ₃	2	H	5.2	6.6352	-0.822
50	CH ₂ C ₆ H ₅	CH ₃	H	2	H	3.7	4.7212	-0.674
51	CH ₂ C ₆ H ₅	H	CH ₃	2	H	0.75	0.957	0.019
52	CH ₂ C ₆ H ₅	H	Cl	2	H	0.17	0.2169	0.664
53	CH ₂ C ₆ H ₅	NO ₂	H	2	H	1.6	2.0416	-0.31
54	CH ₂ C ₆ H ₅	H	NO ₂	2	H	4.8	6.1248	-0.787
55	C ₇ H ₁₅	CH ₃	H	2	H	0.39	0.4976	0.303
56*	C ₇ H ₁₅	H	CH ₃	2	H	0.13	0.1658	0.78
57	C ₇ H ₁₅	H	Cl	2	H	0.013	0.01658	1.78
58*	C ₇ H ₁₅	H	NO ₂	2	H	0.29	0.37	0.431
59	C ₇ H ₁₅	H	OCH ₃	2	H	0.46	0.5869	0.231
60	C ₇ H ₁₅	H	F	2	H	0.045	0.0574	1.241
61(tacrine)	H	H	H	2	H	0.28	0.319	0.498

Table 2

Comp.	R	R1	R2	R3	R4	R5	IC ₅₀ μM	IC ₅₀ #	-log IC ₅₀
62	H	H	H	H	H	H	0.68	0.8636	0.064
63	H	NH ₂	H	H	H	H	5.9	7.493	-0.875
64	H	H	NO ₂	H	H	H	67.5	85.34	-1.931
65	H	H	NH ₂	H	H	H	29	36.83	-1.566
66	H	H	Cl	H	H	H	6.5	8.255	-0.917
67	H	H	F	H	H	H	1.2	1.524	-0.183
68	H	H	H	OCH ₃	H	H	1.6	2.002	-0.308
69	H	H	H	H	OCH ₃	H	6.5	8.255	-0.917
70	H	H	H	H	OCH ₃	H	4.3	5.461	-0.737
71	H	H	H	CH ₃	H	H	3.9	4.953	-0.695
72	H	H	H	H	H	CH ₃	4.6	5.842	-0.766
73	H	H	H	H	F	H	0.43	0.5461	0.263
74	H	H	H	H	Cl	H	5.4	6.858	-0.836
75	CH ₂ C ₆ H ₅	H	H	H	H	H	7.1	9.017	-0.955
76	CH ₂	H	H	H	H	H	1.3	1.651	-0.218
77	C ₇ H ₁₅	H	H	H	H	H	4.3	5.461	-0.737

Normalized IC₅₀ values

Table 3

Comp no	IR2	MRR1	MRR	I ₁	I ₂
1	0	1.03	1.03	1	1
2	0	6.03	1.03	1	1
3	0.71	1.03	1.03	1	1
4	-0.02	1.03	1.03	1	1
5	0.88	1.03	1.03	1	1
6	0.14	1.03	1.03	1	1
7	0	1.03	1.03	1	0
8	0	1.03	5.65	1	1
9	0	1.03	14.96	1	1
10	0	1.03	24.83	1	1
11	0	1.03	34.74	1	1
12	0	1.03	40.2	1	1
13	0	1.03	67.05	1	1
14	0	1.03	41.58	1	1
15	0	1.03	41.58	1	1
16	0	1.03	30.1	1	1
17	0	1.03	35	1	1
18	0	1.03	35	1	1
19	0	1.03	35	1	1
20	0	1.03	29.89	1	1
21	0	1.03	29.89	1	1
22	0	1.03	29.89	1	1
23	0	1.03	36.84	1	1
24	0	1.03	36.84	1	1
25	0	1.03	36.84	1	1
26	0	1.03	34.62	1	1
27	0	1.03	34.62	1	1
28	0	1.03	34.62	1	1
29	0	1.03	33.99	1	1
30	0	1.03	33.99	1	1
31	0	1.03	33.99	1	1
32	0	1.03	25.402	1	1
33	0	1.03	23.54	1	1
34	0.71	1.03	30.1	1	1
35	0.71	1.03	41.88	1	1
36	0.14	1.03	30.1	1	1
37	0.14	1.03	33.99	1	1
38	0.88	1.03	30.1	1	1
39	0	5.65	1.03	0	1
40	0.56	1.03	1.03	0	1
41	0	6.03	1.03	0	1
42	0.71	1.03	1.03	0	1
43	0	7.36	1.03	0	1
44	-0.28	1.03	1.03	0	1
45	-0.02	1.03	1.03	0	1
46	0	5.42	1.03	0	1
47	0.14	1.03	1.03	0	1
48	0.71	6.03	1.03	0	1
49	-0.02	7.87	1.03	0	1
50	0	5.65	30.1	0	1
51	0.56	0	30.1	0	1
52	0.71	1.03	30.1	0	1
53	0	7.36	30.1	0	1
54	-0.28	1.03	30.1	0	1
55	0	5.65	33.53	0	1
56	0.56	1.03	33.53	0	1
57	0.71	1.03	33.53	0	1
58	-0.28	1.03	33.53	0	1
59	-0.02	1.03	33.53	0	1
60	0.14	1.03	33.53	0	1
61	0	1.03	1.03	0	1
62	0	1.03	1.03	0	0
63	0	5.42	1.03	0	0
64	-0.28	1.03	1.03	0	0
65	-1.23	1.03	1.03	0	0
66	0.71	1.03	1.03	0	0
67	0.14	1.03	1.03	0	0
68	0	1.03	1.03	0	0
69	0	1.03	1.03	0	0
70	0	1.03	1.03	0	0
71	0	1.03	1.03	0	0
72	0	1.03	1.03	0	0
73	0	1.03	1.03	0	0
74	0	1.03	1.03	0	0
75	0	1.03	30.1	0	0
76	0	1.03	5.65	0	0
77	0	1.03	33.53	0	0